



**FUNDAÇÃO UNIVERSIDADE FEDERAL DO PAMPA  
PROGRAMA DE PÓS-GRADUAÇÃO EM BIOQUÍMICA**

**INVESTIGAÇÃO DE VIAS DE ESTRESSE OXIDATIVO EM RATOS WISTAR  
HIPERCOLESTEROLÊMICOS SUPLEMENTADOS COM EXTRATO DOS  
FRUTOS DE *Vaccinium ashei* R.**

**DISSERTAÇÃO DE MESTRADO**

**Deise Jaqueline Ströher**

**Uruguaiana, RS, Brasil.**

**2013**

**DEISE JAQUELINE STRÖHER**

**INVESTIGAÇÃO DE VIAS DE ESTRESSE OXIDATIVO EM RATOS WISTAR  
HIPERCOLESTEROLÊMICOS SUPLEMENTADOS COM EXTRATO DOS  
FRUTOS DE *Vaccinium ashei* R.**

Dissertação apresentada ao programa de Pós-graduação *Stricto Sensu* em Bioquímica da Universidade Federal do Pampa, como requisito parcial para obtenção do Título de Mestre em Bioquímica.

Orientadora: Prof. Dr<sup>a</sup>. Vanusa Manfredini

**Uruguiana**

**2013**

**DEISE JAQUELINE STRÖHER**

**INVESTIGAÇÃO DE VIAS DE ESTRESSE OXIDATIVO EM RATOS WISTAR  
HIPERCOLESTEROLÊMICOS SUPLEMENTADOS COM EXTRATO DOS  
FRUTOS DE *Vaccinium ashei* R.**

Dissertação apresentada ao programa de Pós-graduação *Stricto Sensu* em Bioquímica da Universidade Federal do Pampa, como requisito parcial para obtenção do Título de Mestre em Bioquímica.

Área de concentração: Bioprospecção molecular

Dissertação defendida e aprovada em: 22 de outubro de 2013.

Banca examinadora:

---

Prof. Dra. Vanusa Manfredini  
Orientador  
(UNIPAMPA)

---

Prof. Dra. Pâmela Billig Mello Carpes  
(UNIPAMPA)

---

Prof. Dr. Claiton Leoneti Lencina  
(UFPEL)

Estarei contando isso com um suspiro,  
em algum lugar  
daqui a muitas eras.

Duas estradas se bifurcaram numa floresta, e eu...

Eu escolhi a estrada menos percorrida

E isso fez toda a diferença.

Robert Frost

## AGRADECIMENTOS

Gostaria de agradecer a Deus e a Nossa Senhora Aparecida, pela saúde, sabedoria e pela presença constante em minha vida.

Aos meus pais Pedro e Teresinha, pelo apoio e incentivo de sempre. Amo vocês!

Aos meus irmãos Débora e Douglas, por sempre torcerem por mim.

Ao meu namorado, Marcos Raul, pelo incentivo e paciência.

Agradeço com imenso carinho à UNIPAMPA por esta oportunidade, e em especial ao Programa de Pós-Graduação em Bioquímica (PPGBIOQ).

Agradeço aos meus queridos colegas do grupo GESTOX, em especial à Ritiéle, Jamila, Muriel, Maristela, Bruna e Angélica, pela colaboração durante toda a parte experimental deste trabalho. Muito obrigada!

À minha orientadora, Prof. Dra. Vanusa Manfredini pela oportunidade, confiança, compreensão, paciência e amizade. Muito obrigada por tudo!

À Prof. Dra. Jacqueline Piccoli, pela contribuição científica que enriqueceu este trabalho, obrigada!

À Prof. Dra. Francielli Weber Santos e a Aryele pela disponibilidade no auxílio dos experimentos. Obrigada!

À Luisa pela ajuda com o programa de estatística, muito obrigada!

À Ana Eveline pela revisão do inglês para o artigo científico, obrigada!

À CAPES pela concessão da bolsa de estudos.

Aos membros da banca, pela disponibilidade e contribuição científica ao avaliarem este trabalho.

E a todos, que de alguma forma, contribuíram para a concretização deste trabalho, obrigada!

## RESUMO

Dissertação de Mestrado  
Programa de Pós-Graduação em Bioquímica  
Fundação Universidade Federal do Pampa

### INVESTIGAÇÃO DE VIAS DE ESTRESSE OXIDATIVO EM RATOS WISTAR HIPERCOLESTEROLÊMICOS SUPLEMENTADOS COM EXTRATO DOS FRUTOS DE *Vaccinium ashei* R.

AUTORA: Deise Jaqueline Ströher

ORIENTADORA: Vanusa Manfredini

Data e Local da Defesa: Uruguaiana, 22 de outubro de 2013.

A hipercolesterolemia é caracterizada pelo aumento do colesterol total circulante. Uma dieta rica em colesterol aumenta os níveis da lipoproteína de baixa densidade (LDL), que atua como um fator pró-aterogênico, desencadeando processo inflamatório que leva à formação da placa aterosclerótica. Os efeitos dietéticos de plantas sobre o perfil lipídico tem sido documentados e mostram ser úteis na redução dos níveis de colesterol plasmático, prevenindo a aterosclerose. O mirtilo é uma importante fonte alimentar de polifenóis, antocianinas, flavonoides e possui ação antioxidante, conferido o título de alimento funcional. O objetivo do trabalho foi investigar vias de estresse oxidativo em ratos *Wistar* hipercolesterolêmicos suplementados com extrato liofilizado dos frutos de *Vaccinium ashei* R. Os ratos hipercolesterolêmicos foram divididos em 6 grupos ( $n=6$ ): grupo 1: controle (salina); grupo 2: sinvastatina (10mg/Kg) como controle positivo; grupo 3: extrato de mirtilo (25mg/Kg); grupo 4: extrato de mirtilo (50mg/Kg); grupo 5: extrato de mirtilo (25mg/Kg) e sinvastatina (10mg/Kg) e o grupo 6: extrato de mirtilo (50mg/Kg) e sinvastatina (10mg/Kg). Após 14 dias consecutivos de administração, os animais foram eutanasiados e o sangue total e artéria retirados para análises posteriores. O grupo que recebeu o extrato de mirtilo mostrou redução estatisticamente significativa do perfil lipídico (colesterol total, colesterol LDL e triglicerídeos) e um aumento no colesterol HDL. A presença de polifenóis no extrato dos frutos do mirtilo contribuiu para efeito hipolipêmico que ficou mais evidente quando o extrato foi associado à sinvastatina. A dieta hipercolesterolêmica aumentou os níveis séricos da creatina-quinase e homocisteína, porém a administração do extrato diminuiu significativamente os níveis destes marcadores quando comparado ao grupo que não recebeu o extrato. Além disso, o extrato na dose de 50mg/Kg associado à sinvastatina mostrou proteger contra o espessamento da aorta, reduziu a peroxidação lipídica e o dano oxidativo ao DNA induzido por hipercolesterolemia. Os teores de vitamina C e polifenóis séricos aumentaram após 14 dias de administração dos extratos, bem como a atividade das enzimas antioxidantes, superóxido dismutase, catalase e glutathione peroxidase. Assim, os resultados sugerem que o extrato liofilizado de *Vaccinium ashei* R. apresenta um efeito antiaterogênico além de atividade antioxidante e hipocolesterolêmica.

*Palavras chave:* *Vaccinium ashei* Reade, hipercolesterolemia, parâmetros bioquímicos, atividade antioxidante, histologia.

**ABSTRACT**

Dissertation of Master's Degree  
Program of Post-Graduation in Biochemistry  
Federal University of Pampa

**INVESTIGATION OF WAY OF OXIDATIVE STRESS IN RATS WISTAR  
HYPERLIPIDEMICS SUPPLEMENTED WITH EXTRACT OF FRUIT *Vaccinium  
ashei* R.**

AUTHOR: Deise Jaqueline Ströher

ADVISOR: Vanusa Manfredini

Date and Place of Defense: Uruguaiana, October 22<sup>rd</sup>, 2013.

The hypercholesterolemia is characterized by the total circulating cholesterol increase. A diet rich in cholesterol increase the levels of lipoprotein of low density (LDL), that acts as a proatherogenic, triggering inflammatory process that leads to the formation of the atherosclerotic plaque. The dietetic effects of the plants on the lipid profile, have been documented and show to be useful in the plasmatic cholesterol levels reduction, preventing the atherosclerosis. The blueberry fruit is an important source of food rich in polyphenols, anthocyanins, flavonoids, and it has antioxidant action, giving to it the title of functional food. The objective of this study was to investigate ways of oxidative stress in hypercholesterolemic *Wistar* rats, supplemented with lyophilized extract from the *Vaccinium ashei* R fruits. The hypercholesterolemic rats were divided into six groups (n=6): group 1: control (saline); group 2: simvastatin (10mg/Kg) as positive control; group 3: blueberry extract (25mg/Kg); group 4: blueberry extract (50mg/Kg); group 5: blueberry extract (25mg/Kg) and simvastatin and the group 6: blueberry extract (50mg/Kg) and simvastatin. After 14 consecutive days of administration, the animals were euthanized and the total blood and arteries were removed to be analyzed later. The group that received the blueberry extract shown statistically significant reduction of the lipid profile (total cholesterol, LDL-cholesterol and triglycerides) and an increase of the HDL-cholesterol. The presence of polyphenols in the blueberry extract contributed to hypolipidemic effect what is more evident when the extract is associated with to the simvastatin. The hypercholesterolemic diet increased the serum levels of creatine kinase and homocysteine, nevertheless, the administration of the extract decrease significantly the levels of these makers when they are compared to the group that did not received the extract. Moreover, the extract at a dose of 50mg/kg associated with simvastatin showed protection against the aorta thickening, it reduced the lipid peroxidation and oxidative damage to DNA, induced by hypercholesterolemia. The contents of vitamin C and serum polyphenols increased after 14 days of the extracts administration, as the activity of the oxidant enzymes, superoxide dismutase, catalase and glutathione peroxidase. In this manner, the results point that the lyophilized extract of *Vaccinium ashei* R. shows an antiatherogenic effect, besides the antioxidant and hypocholesterolemic activities.

**Keywords:** *Vaccinium ashei* Reade, hypercholesterolemia, biochemical parameters, antioxidant activity, histology.

## LISTA DE ILUSTRAÇÕES

<b>Figura 01</b> – Etapas do processo de formação da placa aterosclerótica.....	18
<b>Figura 02</b> - <i>Vaccinium ashei</i> Reade.....	27
<b>Figura 03</b> - Estrutura geral dos flavonoides.....	29
<b>Figura 04</b> - Estrutura geral das diferentes subclasses de flavonoides.....	29
<b>Figura 05</b> - Estruturas das antocianinas mais comumente encontradas na natureza.....	30

### Manuscrito I

<b>Figure 01</b> - Effect of blueberry extract in hypercholesterolemic rats after treatment.....	61
<b>Figure 02</b> - Histological section and thickening measure of the aorta of hypercholesterolemic rats after treatment.....	64

### Manuscrito II

<b>Figure 01</b> - Antioxidant activities <i>in vitro</i> of the lyophilized blueberry extract of <i>Vaccinium ashei</i> R determined at different concentrations: (A) DPPH radical-scavenging activity. (B) ABTS radical-scavenging activity.....	88
<b>Figure 02</b> - Oxidative damage markers in hypercholesterolemic rats after treatment of blueberry extract.....	89
<b>Figure 03</b> - Antioxidative defenses markers in in hypercholesterolemic rats after treatment of blueberry extract.....	90



**LISTA DE TABELAS****Manuscrito 1**

<b>Table 01</b> - Body weight of rats during the induction of hypercholesterolemia (weeks 1-4) and during treatment (weeks 5-6).....	60
<b>Table 02</b> - Hematological parameters of the hyperlipidemic rats exposed to different treatments.....	62
<b>Table 03</b> – Markers of cardiac damage in the hypercholesterolemic rats exposed to different treatments.....	63

## LISTA DE ABREVIATURAS

- AF** – Alimentos funcionais
- ANVISA** – Agência Nacional de Vigilância Sanitária
- CAT** – Catalase
- CEP** - Comitê de ética em pesquisa
- DCV** – Doenças Cardiovasculares
- DNA** – Ácido desoxirribonucleico
- DTNB** - Ácido 5,5'-ditio-bis(2-nitrobenzóico)
- ERN** – Espécies reativas de nitrogênio
- ERO** – Espécies reativas de oxigênio
- GPx** – Glutathione peroxidase
- GSH** – Glutathione reductase
- H<sub>2</sub>O<sub>2</sub>** - Peróxido de hidrogênio
- HDL**- Lipoproteína de alta densidade
- HOCl** – Ácido hipocloroso
- LDL** – Lipoproteína de baixa densidade
- LDL ox** – Lipoproteína de baixa densidade oxidada
- MCP-1** – Proteína quimiotática para monócitos 1
- M-CSF** – Fator estimulante de colônias de monócitos
- MM-LDL-OX** – Lipoproteína de baixa densidade minimamente oxidada
- MS** - Ministério da Saúde
- N<sub>2</sub>O** – Óxido nitroso
- NO<sup>•</sup>** - Óxido nítrico
- O<sub>2</sub>** - Oxigênio singleto
- O<sub>2</sub><sup>•-</sup>** - Ânion superóxido
- OH<sup>•</sup>** - Radical hidroxila
- ONOO<sup>-</sup>** - Peroxinitrito
- RL** – Radicais livres
- RO<sup>•</sup>** - Radical alcóxila
- ROO<sup>•</sup>** - Radical peróxila
- SOD** – Superóxido dismutase
- TBARS** - Espécies reativas ao ácido tiobarbitúrico

## SUMÁRIO

<b>PARTE I</b> .....	15
<b>1.0 INTRODUÇÃO</b> .....	15
<b>2.0 REVISÃO BIBLIOGRÁFICA</b> .....	17
2.1 Doenças Cardiovasculares e Aterosclerose .....	17
2.2 Radicais livres, Estresse Oxidativo e Defesas Antioxidantes.....	21
2.3 Alimentos Funcionais .....	24
2.4 Mirtilo .....	26
<b>3.0 OBJETIVOS</b> .....	33
3.1 Objetivo geral: .....	33
3.2 Objetivos específicos:.....	33
<b>PARTE II</b> .....	34
<b>MANUSCRITO I</b> .....	34
1.0 INTRODUCTION .....	40
2.0 MATERIALS AND METHODS .....	41
2.1 Sample plant .....	41
2.2 Preparation of the extract.....	41
2.3 Animal Experimentation .....	42
2.4 Preparation of feed and Induction of hypercholesterolemia.....	42
2.5 Experimental Desing .....	43
2.6 Blueberry extract prepare and administration .....	43
2.7 Biochemical, hematological and cardiac markers analysis .....	43
2.8 Collection of Aorta and Histopathological analysis .....	44
2.9 Statistical Analysis .....	44
3.0 RESULTS .....	44
3.1 Effects of BE on body weight gain.....	44

3.2 Effect of the BE treatment on Biochemical Profile .....	45
3.3 Effect of the BE treatment on Hematological Profile.....	46
3.4 Effect of the BE treatment on Markers of cardiac damage .....	46
3.5 Effect of the BE treatment on Histopathological Profile.....	47
4.0 DISCUSSION.....	47
5.0 LITERATURE CITED.....	52
<b>MANUSCRITO II</b> .....	<b>65</b>
1.0 INTRODUCTION .....	68
2.0 MATERIALS AND METHODS .....	69
2.1 Chemicals .....	69
2.2 Plant Material .....	69
2.3 Preparation of the extract.....	69
2.4 Evaluation of antioxidant potential from blueberry extract in vitro .....	69
2.5 Evaluation of blueberry extract in vivo .....	71
2.6 Evaluation of oxidative stress parameters .....	72
2.7 Statistical Analysis .....	73
3.0 RESULTS/DISCUSSION .....	73
3.1 Radical scavenging capacity of lyophilized blueberry extract .....	73
3.2 The total polyphenols content.....	74
4.1 Evaluation of oxidative stress parameters .....	74
4.0 REFERENCES .....	81
<b>PARTE III</b> .....	<b>91</b>
<b>4.0 CONCLUSÃO</b> .....	<b>91</b>
<b>5.0 PERSPECTIVAS</b> .....	<b>92</b>
<b>6.0 REFERÊNCIAS BIBLIOGRÁFICAS</b> .....	<b>93</b>
<b>7.0 ANEXOS</b> .....	<b>102</b>
Protocolo de aprovação do projeto pelo CEUA-UNIPAMPA .....	102

Certificado de apresentação do trabalho..... 103

## APRESENTAÇÃO

A presente dissertação foi dividida em três partes principais. Na **parte I** encontram-se a **INTRODUÇÃO**, **REFERENCIAL TEÓRICO** e **OBJETIVOS**. Os resultados que fazem parte desta dissertação estão apresentados sob a forma de manuscritos, os quais se encontram no item **MANUSCRITOS**, **parte II** deste trabalho. As seções materiais e métodos, resultados, discussão dos resultados e referências bibliográficas, encontram-se nos próprios manuscritos e representam a íntegra deste estudo. O item **CONCLUSÃO** encontra-se na **parte III** desta dissertação, apresenta interpretações e comentários gerais sobre os resultados apresentados nos manuscritos deste trabalho. O item **REFERÊNCIAS BIBLIOGRÁFICAS** refere-se somente às citações que aparecem nos itens introdução, referencial teórico e conclusão desta dissertação. No item **PERSPECTIVAS**, estão expostos os possíveis estudos para dar continuidade a este trabalho.

## PARTE I

### 1.0 INTRODUÇÃO

As doenças cardiovasculares (DCV) são a maior causa de morbimortalidade, tanto em países desenvolvidos quanto em países em desenvolvimento. Os principais fatores de risco associados ao desenvolvimento das DCV são o tabagismo, obesidade e dislipidemia (SPOSITO *et al.*, 2007).

A dislipidemia constitui o maior fator de impacto no desenvolvimento da doença aterosclerótica, em particular a presença de concentrações aumentadas de lipoproteína de baixa densidade (LDL) (SPOSITO *et al.*, 2007). Nesse sentido a hipercolesterolemia mostra-se um fator muito importante, pois este evento pode desencadear outras patologias, como a aterosclerose, infarto do miocárdio, acidente vascular cerebral, diabete mellitus e outras doenças renais e hepáticas (SCHIAVO *et al.*, 2003).

O estresse oxidativo ocupa um local de destaque nas pesquisas com aterosclerose, visto que a modificação oxidativa da LDL é a hipótese mais referendada em ser responsável para o início e a progressão do processo aterosclerótico (ROSENSON, 2004). Ele ocorre quando há um desequilíbrio no estado redox do organismo, gerando um excesso de radicais livres (RL), que são capazes de danificar biomoléculas como proteínas citosólicas, lipídeos de membrana e o ácido desoxirribonucleico (DNA) (SPEIT *et al.*, 1996; HALLIWELL & GUTTERIDGE, 2007).

A fim de contrapor este quadro o organismo humano possui mecanismos de defesa antioxidante, que atuam intracelular e extracelularmente para manter o equilíbrio redox da célula, assegurando que o aumento das espécies reativas de oxigênio (ERO) seja transitório (RIBEIRO *et al.*, 2008).

A utilização de plantas medicinais como recurso terapêutico é uma tendência milenar, que se encontra em plena ascensão, contribuindo significativamente para sanar as necessidades primárias de assistência à saúde. Aproximadamente 80% da população mundial utiliza a medicina tradicional, fato que se deve a cultura ou a falta de

alternativas (WHO, 2011). No Brasil, além do uso de plantas medicinais, a utilização de alimentos funcionais (AF) encontra-se em expansão e contribui significativamente para auxiliar a promoção da saúde. Os AF são ingredientes que produzem efeitos metabólicos e/ou fisiológicos e/ou efeitos benéficos à saúde, além de suas funções nutricionais básicas. Este efeito ocorre em sua maioria quando estes são consumidos como parte da dieta (ERLUND *et al.*, 2003).

As frutas do gênero *Vaccinium* podem ser consideradas como os primeiros AF descritos. O mirtilo é uma espécie frutífera originária de algumas regiões da Europa e América do Norte, onde é muito apreciado por seu sabor exótico, pelo valor econômico e por seus poderes medicinais, sendo considerada como “fonte de longevidade”. Além disso, possui um alto conteúdo de polifenóis, como os flavonóides e antocianidinas, com poder antioxidante contido nos pigmentos de cor azul-púrpura dos frutos (BASU *et al.*, 2010; WU *et al.*, 2010).

Estudos sugerem que os frutos mirtilo apresentam muitos benefícios à saúde, tais como atividade antioxidante, atividade antidiabética (LEDUC *et al.*, 2006) e capacidade de proteger contra o acidente vascular cerebral e o câncer (WANG *et al.*, 2005). Além disso, a ingestão do extrato de frutos de mirtilo tem mostrado melhorar a memória de curto prazo, o equilíbrio e a coordenação em ratos velhos (JOSEPH, *et al.*, 1999).

Assim, considerando o importante papel dos elevados níveis plasmáticos dos lipídeos no desenvolvimento da resistência a insulina, aterosclerose e DCV (BARBALHO *et al.*, 2009), este trabalho teve como objetivo investigar vias de estresse oxidativo em ratos *Wistar* hipercolesterolêmicos suplementados com extrato dos frutos de *Vaccinium ashei* Reade.



## 2.0 REVISÃO BIBLIOGRÁFICA

### 2.1 Doenças Cardiovasculares e Aterosclerose

Dietas ricas em colesterol, bem como a modificação dos padrões dietéticos que incluem o aumento no consumo de energia, açúcares, sal, gorduras totais, trans e saturadas (MAGALHÃES, CHAGAS e LUZ, 2005) aliadas à inatividade física, possibilitam o aumento na prevalência de dislipidemia, que é considerada um dos principais fatores de risco para DCV. As DCV e suas complicações permanecem a principal causa de morte em países industrializados (LOPEZ *et al.*, 2006).

A DCV é caracterizada pela elevação nos níveis plasmáticos de triacilgliceróis, colesterol total e sua fração LDL, associados à diminuição dos valores da lipoproteína de alta densidade (HDL) (BRUCKNER, 2008). Nos últimos anos, o uso do grupo de fármacos denominado estatinas, foi estabelecido como uma terapia eficaz para reduzir os níveis de LDL e, conseqüentemente, o risco cardiovascular (BAIGENT *et al.*, 2010).

O desenvolvimento das DCV é influenciada por diferentes fatores que incluem hipertensão arterial, hipercolesterolemia, tabagismo, diabetes mellitus, obesidade, herança genética, sedentarismo e o estresse (KAWAMORI *et al.*, 1992; BECKSTROM *et al.*, 2007). Além destes, homocisteinemia aumentada e função plaquetária alterada também são citados como fatores que predispõem ao desenvolvimento da placa de ateroma (PICCINATO; CHERRI; MORIYA, 2001).

Durante a última década, muitos trabalhos tem mostrado informações detalhadas sobre os acontecimentos inflamatórios que ocorrem na aterosclerose. No entanto, a maneira pela qual esses eventos contribuem para a formação, evolução e complicação de lesões ainda não é completamente entendido. Diferentes autores tem estabelecido ligações entre a hipercolesterolemia e o processo inflamatório que ocorre na aterosclerose. Porém, sabe-se que as LDL, quando retidas e oxidadas na camada íntima da artéria, são as principais indutoras da ativação inflamatória da célula endotelial, que leva ao início e a progressão do processo aterosclerótico (STEINBERG *et al.*, 2009; NAVAB *et al.*, 2004).

A oxidação da LDL ocorre em pequena proporção ainda na circulação sanguínea e continua após a entrada da LDL na camada íntima das artérias, em ambiente pró-oxidante (KOVANEN e PENTIKAINEN, 2003). As partículas responsáveis por esta oxidação da LDL são os radicais livres (RL) que podem ser as espécies reativas de oxigênio (ERO) e as espécies reativas de nitrogênio (ERN).

As partículas de LDL difundem-se passivamente através das células endoteliais por transporte vesicular, o qual não necessita de receptores, e aderem à parede do vaso por interações entre a apoproteína B, presente na sua estrutura, e os proteoglicanos da matriz subendotelial (Figura 1) (LUSIS, 2000).

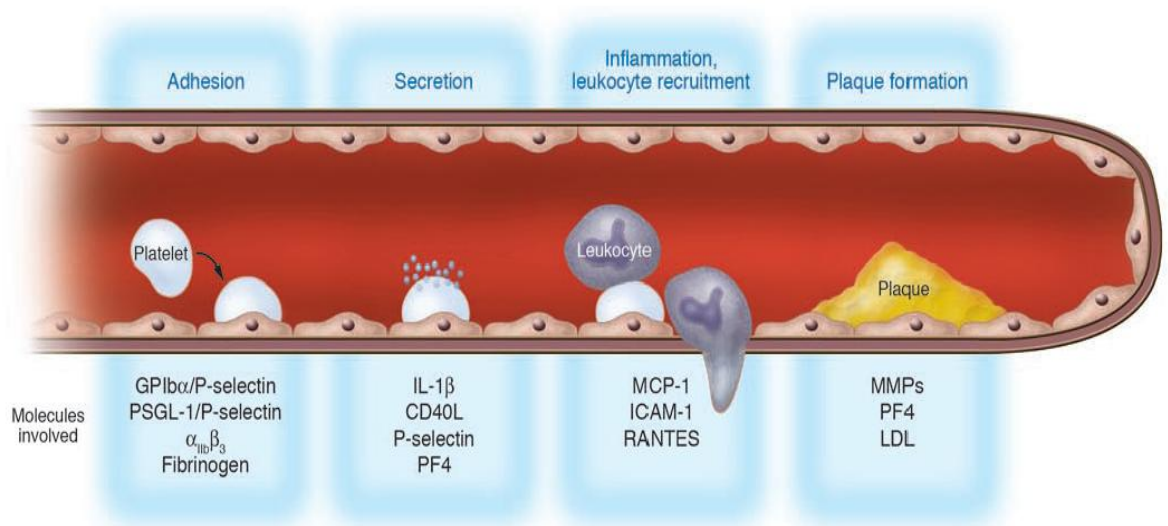


Figura 1 – Etapas do processo de formação da placa aterosclerótica  
(Adaptado de GAWAZ *et al.*, 2005).

Retidas na camada íntima da artéria, a LDL sofre modificações oxidativas, que parecem ocorrer em dois estágios. O primeiro, antes que os monócitos sejam ativados, resulta na oxidação dos lipídios da LDL, com pequena alteração na apoproteína B, resultando na LDL minimamente oxidada (MM-LDL-OX). A MMLDL-OX estimula as células endoteliais a produzirem moléculas aterogênicas e pró-inflamatórias, como a proteína quimiotática para monócitos 1 (MCP-1), que promove a quimiotaxia de monócitos e linfócitos T para o espaço subendotelial; moléculas de adesão e o fator

estimulante de colônias de monócitos (M-CSF), que ativa a diferenciação de monócitos em macrófagos (ROSS, 1999).

O segundo estágio de oxidação da LDL, formando a LDL oxidada (LDLox), ocorre quando monócitos são recrutados para a lesão e convertidos em macrófagos, contribuindo com sua enorme capacidade oxidativa. Nesta fase, os lipídios da LDL são adicionalmente oxidados e a parte protéica também é modificada, impedindo o reconhecimento da lipoproteína pelo receptor de LDL, tornando-a reconhecível apenas pelos receptores presentes nos macrófagos (BROWN e GOLDSTEIN, 1990).

Dentro dos macrófagos, a LDLox é degradada, o colesterol livre é esterificado, conferindo às células o aspecto de espuma. O resultado é o grande acúmulo de colesterol e a formação de células espumosas, originando a primeira lesão da aterosclerose: a estria gordurosa (STEINBERG, 1997).

Posteriormente, as células musculares lisas começam a migrar da camada média da parede arterial para a íntima ou espaço subendotelial, se proliferam e secretam colágeno, dando origem à lesão intermediária. Nesta fase, o espessamento da íntima provoca o remodelamento, ou seja, uma dilatação da artéria compensatória ao estreitamento do lúmen. Mais uma vez, sob o estímulo da LDLox, entre outros, as células do sistema imune local liberam enzimas, citocinas e fatores de crescimento que podem induzir necrose. Ciclos repetidos de acúmulo e ativação de células mononucleares, migração e proliferação das células musculares lisas com produção de colágeno, levam ao aumento progressivo da lesão, até que se estruture uma capa fibrosa ao redor de um núcleo lipídico e de tecido necrótico, a chamada lesão avançada (ROSS, 1999).

As lesões avançadas estáveis são mais resistentes à ruptura e se caracterizam por células musculares lisas envoltas de densa matriz de colágeno, com baixo conteúdo de células inflamatórias e de lipídios no centro necrótico. Por outro lado, as regiões onde as lesões apresentam volumoso centro necrótico e um grande infiltrado de células espumosas, capa fibrosa frágil e fina, com pouca quantidade de colágeno, são mais susceptíveis à ruptura, caracterizando lesões instáveis (LEE e LIBBY, 1997).

A ruptura da capa fibrosa expõe material lipídico altamente trombogênico, levando à formação de um trombo sobrejacente. Este processo, também conhecido por

aterotrombose, é um dos principais determinantes das manifestações clínicas da aterosclerose (SPOSITO *et al.*, 2007).

A LDLox participa de todas as etapas do processo de aterosclerose, desde a disfunção endotelial até a formação das placas ateroscleróticas (SIQUEIRA *et al.*, 2006). Produtos derivados da LDLox são citotóxicos, podendo promover a apoptose celular. Em adição, a LDLox também contribui para o processo inflamatório na aterosclerose por inibir a produção de óxido nítrico (NO), que é um vasodilatador, e por estimular a produção de citocinas, como a interleucina-1, e aumentar a agregação plaquetária (SINGH e JIALAL, 2006).

## 2.2 Radicais livres, Estresse Oxidativo e Defesas Antioxidantes

O termo radical livre (RL) é frequentemente usado para designar qualquer átomo ou molécula contendo um ou mais elétrons desemparelhados nos orbitais mais externos, o que torna essas moléculas altamente reativas e capazes de reagir com qualquer composto que esteja próximo, passando a assumir uma ação oxidante (HALLIWELL e GUTTERIDGE, 1999).

Entretanto, o termo não é ideal para designar todos os agentes reativos, pois, alguns destes não possuem elétrons desemparelhados, como é o caso do peróxido de hidrogênio ( $H_2O_2$ ). Apesar de o  $H_2O_2$  não ser um RL, ele pode ser bastante danoso às células, principalmente devido à reação entre ele e o ânion superóxido, formando o radical hidroxila ( $OH\bullet$ ), altamente reativo (HALLIWELL, 1991).

Os RL cujo elétron encontra-se centrado nos átomos de oxigênio ou nitrogênio são denominados, respectivamente, de ERO e ERN (ABRAHÃO, 2007). As principais ERO distribuem-se em dois grupos, as radicalares: ânion superóxido ( $O_2^{\bullet-}$ ), radical hidroxila ( $OH\bullet$ ), peroxila ( $ROO\bullet$ ) e alcoxila ( $RO\bullet$ ); e as não radicalares: oxigênio singlete ( $O_2$ ), peróxido de hidrogênio ( $H_2O_2$ ) e ácido hipocloroso ( $HOCl$ ). Dentre as ERN incluem-se óxido nítrico ( $NO\bullet$ ), óxido nitroso ( $N_2O$ ) e peroxinitrito ( $ONOO^-$ ), dentre outros (GILLHAM *et al.*, 1997).

Em condições fisiológicas normais, as ERO possuem um papel importante em seres vivos, como a regulação da resposta imunológica, participando do processo fagocítico de defesa contra infecções e atuando como fatores de transcrição na sinalização intracelular, induzindo apoptose (HALLIWELL, 1994; BIESALSKI, 2002). No entanto, em determinadas condições, pode ocorrer um aumento na produção de ERO e/ou a redução na sua eliminação pelas defesas antioxidantes, causando um desequilíbrio fisiológico, que resulta no chamado estresse oxidativo (FINKEL e HOLBROOK, 2000; GUTTERIDGE e HALLIWELL, 2000).

O desequilíbrio entre a produção e a remoção das ERO, podem ocasionar vários eventos nocivos, como a apoptose de células saudáveis, o envelhecimento precoce, a alteração da função celular e o aparecimento de doenças degenerativas como aterosclerose, câncer, doença de Alzheimer ou a doença de Parkinson (FINAUD, *et al.*, 2006). Além disso, o estresse oxidativo está envolvido na oxidação das LDL e tem

demonstrado ser um importante fator no desenvolvimento de doenças cardiovasculares (MAYNE, 2003).

Um antioxidante pode ser definido como uma substância que, em baixa concentração em relação a um determinado substrato, retarda ou previne a oxidação do substrato oxidável (HALLIWEL *et al.*, 1995). Quando o mecanismo de ação for através de sua reação com os RL, o novo radical formado deve ser estável e incapaz de propagar a reação (SHAHIDI *et al.*, 1992).

Para combater os danos deletérios causados pelo estresse oxidativo, o organismo humano possui mecanismos de defesa antioxidante, os quais atuam intracelular e extracelularmente e mantêm o equilíbrio redox da célula, assegurando que o aumento das ERO seja transitório. Existem dois mecanismos antioxidantes: o enzimático e o não enzimático, os quais agem cooperativamente para manter o equilíbrio dos RL no organismo, e, em consequência disso, diminuir o dano às estruturas biológicas (RIBEIRO *et al.*, 2008; BELLÓ, 2002).

O mecanismo de defesa enzimático é a primeira linha de defesa do organismo contra os danos oxidativos. O sistema é constituído por um conjunto de enzimas, tais como a superóxido dismutase (SOD), glutatona peroxidase (GPx), glutatona redutase (GSH), catalase (CAT), tioredoxinas, peroxirredoxinas e inúmeras outras redutases (RIBEIRO *et al.*, 2008). Já o mecanismo não-enzimático é constituído por um grande número de compostos de baixo peso molecular, ingeridos pela dieta (nutrientes e não-nutrientes) como as vitaminas A, C, E e os flavonoides, ou sintetizados no organismo, como a glutatona (RIBEIRO *et al.*, 2008; MANACH *et al.*, 2004).

Quando a produção de RL e/ou espécies reativas supera a capacidade de ação dos antioxidantes, ocorre a oxidação de biomoléculas, gerando metabólitos específicos que são os marcadores do estresse oxidativo. Tais marcadores são derivados, sobretudo, da oxidação de lipídeos, proteínas e DNA (HALLIWEL e WHITEMAN, 2004; VINCENT, *et al.*, 2007; MAYNE, 2003).

Nos últimos anos, tem aumentado a busca por compostos naturais eficazes, não tóxicos, com atividade antioxidante e que possam ser utilizados na prevenção e tratamento de doenças. Tentativas de utilização de antioxidantes sintéticos para bloquear ou atenuar os efeitos prejudiciais de ERO têm produzido resultados negativos (COZMA, 2004; CHEN, *et al.*, 2005; PAPA HARALAMBUS *et al.*, 2007), e cada vez

mais atenção tem sido dada aos produtos naturais (BANDYOPADHYAY, *et al.*, 2004). Nesse contexto, inserem-se os AF, considerados promotores de saúde por estarem associados à diminuição dos riscos de doenças crônicas.

### 2.3 Alimentos Funcionais

Ainda não existe um consenso mundial a respeito do que são os AF, no entanto, a definição mais comum indica que um alimento pode ser considerado funcional se for demonstrado que pode influenciar positivamente uma ou mais funções alvo no corpo, além de possuir efeitos nutricionais adequados, de maneira a ser tanto relevante para o bem-estar e a saúde, quanto para a redução do risco de doença (ROBERFROID, 2002).

Os AF representam um conceito, mais do que um grupo definido de alimentos, porém a área é precisamente a de alimentos. Eles possuem um valor nutricional, aspecto, propriedades sensoriais e demais atributos de todo alimento, porém, não são um veículo de fármacos com ação farmacológica. Representam uma conjunção com princípios ativos, que quando consumidos em uma quantidade razoável exercem ações benéficas à nível fisiológico (NOONAN e NOONAN, 2004).

No Brasil, o Ministério da Saúde (MS), através da Agência Nacional de Vigilância Sanitária (ANVISA), regulamentou os AF através das seguintes resoluções: ANVISA/MS 16/99; ANVISA/MS 17/99; ANVISA/MS 19/99. Segundo a ANVISA, AF são aqueles que produzem efeitos metabólicos ou fisiológicos através da atuação de um nutriente ou não-nutriente no crescimento, desenvolvimento, manutenção e em outras funções normais do organismo humano.

Para Lajolo (2001), “alimento funcional é o alimento semelhante em aparência ao alimento convencional, consumido como parte da dieta usual, capaz de produzir demonstrados efeitos metabólicos e fisiológicos úteis na manutenção de uma boa saúde física e mental, podendo auxiliar na redução do risco de doenças crônico-degenerativas, além das suas funções nutricionais básicas”.

De acordo com Pimentel e colaboradores (2005) os AF são classificados baseados na sua natureza química e molecular. Para eles os alimentos funcionais são classificados em sete grupos, sendo eles: isoprenóides, compostos fenólicos, proteínas, carboidratos e derivados, ácidos graxos e lipídeos, minerais e os microbióticos.

Em muitas partes do mundo os AF e os nutracêuticos possuem conceituações semelhantes, porém há uma diferença fundamental entre eles, que faz com que os AF se relacionem à venda e consumo dos mesmos como alimentos, ao passo que os



nutracêuticos são ingredientes funcionais isolados e podem ser consumidos sob diferentes formas, dadas pela indústria farmacêutica.

O Brasil é um país rico em produtos naturais e AF ainda não explorados (CUPPARI, 2002). Aliado a isso, o aumento da expectativa de vida da população e o crescimento exponencial de doenças crônicas tais como a obesidade, a aterosclerose, a hipertensão, o diabetes e o câncer têm ocasionado um crescente interesse na busca por novos AF capazes de prevenir ou reduzir o risco de ocorrência dessas e outras patologias.

## 2.4 Mirtilo

### 2.4.1 Características gerais e produção

O mirtilo é a fruta do mirtilheiro, planta frutífera de clima temperado que é membro da família *Ericaceae* e pertence ao gênero *Vaccinium* (RASEIRA E ANTUNES, 2004), originária de algumas regiões da Europa e América do Norte, onde é conhecida popularmente como “blueberry”. O mirtilo é um fruto tipo baga, possui uma coloração azul-escura e um sabor agridoce (KLUGE *et al.*, 1994) e, é muito apreciado por seu sabor exótico, pelo valor econômico e por seus poderes medicinais, sendo considerado como fruto “fonte da longevidade” (ANTUNES; MADAIL, 2005).

A produção de mirtilo está concentrada principalmente nos Estados Unidos e no Canadá, onde, o primeiro, é responsável por 66% e o segundo por 33% da produção mundial (STRIK, 2005). Os trabalhos com o mirtilo no Brasil iniciaram em 1983, na Embrapa Clima Temperado (Pelotas-RS), com a introdução da coleção de cultivares de baixa exigência em frio do grupo “*rabbiteye*” (SILVA *et al.*, 2008), espécie considerada pelos produtores como a que oferece as maiores possibilidades para adaptação (ECK *et al.*, 1990).

O crescente interesse pelas frutas tem mobilizado o mercado mundial a aumentar a oferta do fruto, expandindo seu cultivo em países da América do Sul, como Chile, Argentina e Uruguai (BAÑADOS, 2006; RASEIRA e ANTUNES, 2004). O Brasil é um produtor ainda recente de mirtilo (FACHINELLO, 2008) e o potencial cultivo do fruto, aponta o *Vaccinium ashei* R como a espécie mais adaptável às condições de clima frio do Sul do Brasil (Figura 2).



Figura 2 - *Vaccinium ashei* Reade

Fonte: Embrapa

Atualmente, a produção se concentra nas regiões sul e sudeste do país, nos municípios Vacaria e Caxias do Sul (Rio Grande do Sul, RS), Barbacena (Minas Gerais, MG), e Campos do Jordão (São Paulo, SP) (SANTOS, 2004), porém, o estado do Rio Grande do Sul ainda é o que mais se destaca na produção de mirtilo (KLUGE *et al.*, 1994).

Segundo Raseira e Antunes (2004), existem muitas espécies de mirtilo, sendo que as principais espécies com expressão comercial são divididas em três grupos, de acordo com o genótipo, hábito de crescimento, tipo de fruto produzido e outras características. Estes grupos são:

- a) “highbush”: sua produção, dentre os demais grupos, é a de melhor qualidade, tanto em tamanho quanto em sabor dos frutos. A principal espécie deste grupo é *Vaccinium corymbosum* L.;
- b) “rabbiteye”: compreende a espécie *Vaccinium ashei* Reade. Em relação ao grupo anterior, produz frutos de menor tamanho e de menor qualidade. Apresenta maior produção por planta, e seus frutos têm maior conservação em pós-colheita. Apresenta maior importância comercial em regiões com menor disponibilidade de frio, por causa da sua tolerância a temperaturas mais elevadas e à deficiência hídrica;
- c) “lowbush”: tem hábito de crescimento rasteiro e produz frutos de pequeno tamanho, cujo destino é a indústria processadora.

### 2.4.2 Compostos fenólicos

Os vegetais produzem uma grande variedade de substâncias que não possuem ação direta na fotossíntese, respiração, síntese de proteínas, de carboidratos e de lipídeos, sendo considerados compostos produzidos pelo metabolismo secundário (TAIZ & ZEIGER, 2004). Os compostos fenólicos ou polifenóis fazem parte do metabolismo secundário vegetal e possuem diversas funções nos vegetais, tais como proteção contra raios ultravioleta, proteção contra insetos e bactérias, controle da ação de hormônios vegetais, além de atrair animais com finalidade de polinização (ZUANAZZI e MONTANHA, 2004).

Quimicamente, os compostos fenólicos podem ser definidos como substâncias que possuem um anel aromático contendo um ou mais substituintes hidroxila (LEE *et al.*, 2005) e a atividade antioxidante destes compostos depende da sua estrutura, particularmente do número e posição dos grupos hidroxila e da natureza das substituições nos anéis aromáticos (BALASUNDRAM, SUNDRAM e SAMMAN, 2006).

O mirtilo (*Vaccinium* sp.) apresenta em sua composição alta concentração de compostos fenólicos (WU *et al.*, 2004), sendo superior a maioria das frutas (WOLFE *et al.*, 2008). No entanto, estudos demonstram que há uma grande variação qualitativa e quantitativa na composição fenólica do mirtilo e que, esta variação é dependente de fatores intrínsecos (gênero, espécie e cultivar) e extrínsecos (condições ambientais, cultivo, manejo e condições de armazenamento) (WANG *et al.*, 2008; GIOVANELLI e BURATTI, 2009).

### 2.4.3 Flavonóides

Os flavonóides constituem o maior grupo dos compostos fenólicos, sendo descritos mais de 8.000 compostos (BEECHER, 2003). Estes são pigmentos responsáveis pelas cores amarelas, laranjas e vermelhas das flores, sendo importantes para o desenvolvimento e para defesa das plantas (RICE-EVANS, 2003). Estes compostos possuem uma estrutura comum de difenilpropanos (C6 – C3 – C6), constituídos de dois anéis aromáticos e um heterociclo oxigenado ligados através de três

carbonos (Figura 3), os quais se subdividem em seis subclasses como isoflavona, antocianina, flavanona, catequina, flavona e flavonol (ROSS & KASUM, 2002) representadas na Figura 4.

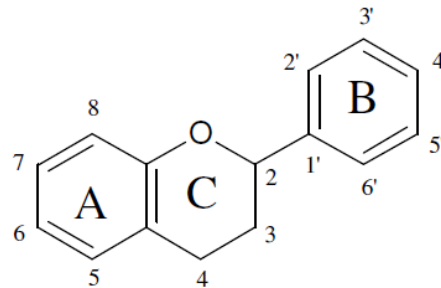


Figura 3 - Estrutura geral dos flavonóides

(Adaptado de: BALASUNDRAM, SUNDRAM e SAMMAN, 2006).

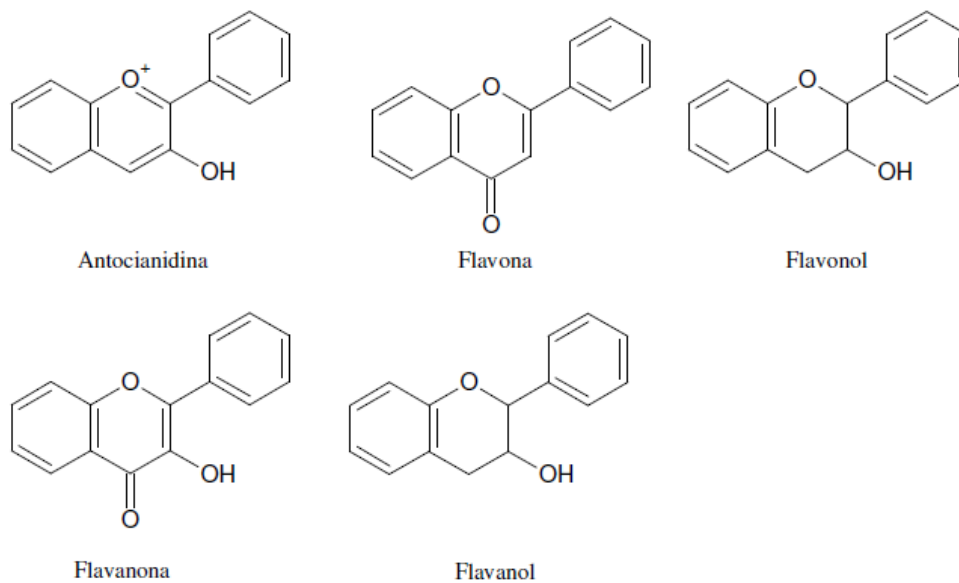


Figura 4 - Estrutura geral das diferentes subclasses de flavonóides

(Adaptado de: BALASUNDRAM, SUNDRAM e SAMMAN, 2006).

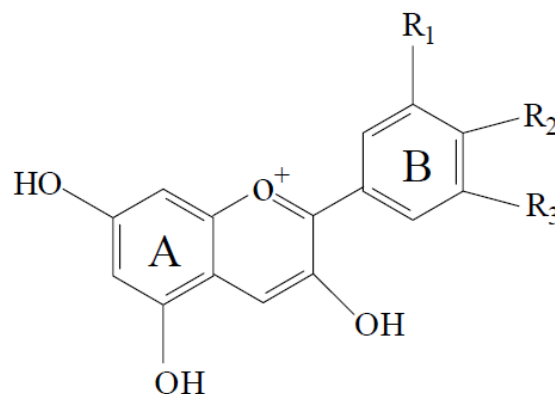
#### 2.4.4 Antocianinas

As antocianinas (do grego: *anthos*, flor e *kyanos*, azul) pertencem a uma das subclasses dos flavonóides, sendo responsáveis pela coloração atraente de diversas

flores e frutas. Seu espectro de cor pode variar de salmão, rosa, vermelho, magenta, violeta, roxo e azul (MANHITA, *et al.*, 2006; CISSE *et al.*, 2009).

A estrutura das antocianinas é constituída por dois anéis benzênicos e um anel heterocíclico central, contendo oxigênio (KONCZAK, 2004). Essa estrutura é completada por uma ou mais moléculas de açúcar ligadas em diferentes posições hidroxiladas da estrutura básica (DELGADO-VARGAS & PAREDES-LÓPEZ, 2003) (Figura 5).

As antocianinas diferem entre si pelo número de grupos hidroxila, número e natureza dos açúcares unidos à molécula, posição desse açúcar e pelo número e natureza dos ácidos alifáticos ou aromáticos unidos aos açúcares da molécula (KONG *et al.*, 2003) e, devido à sua polaridade, são mais solúveis em solventes polares que em apolares (DELGADO-VARGAS & PAREDES-LÓPEZ, 2003). A Figura 5 ilustra a estrutura de uma antocianina.



Antocianinas	R <sub>1</sub>	R <sub>2</sub>	R <sub>3</sub>
Cianidina	OH	OH	-
Peonidina	OCH <sub>3</sub>	OH	-
Delfinidina	OH	OH	OH
Malvidina	OCH <sub>3</sub>	OH	OCH <sub>3</sub>
Petunidina	OCH <sub>3</sub>	OH	OH

Figura 5 - Estruturas das antocianinas mais comumente encontradas na natureza.

(Adaptado de: JACKSON, 1994).

As antocianinas são glicosídeos que apresentam em sua estrutura química um resíduo de açúcar na posição 3, facilmente hidrolizado, como produtos desta hidrólise

obtem-se o componente glicídico e a aglicona, denominadas antocianidina (DEWICK, 2002).

Aproximadamente 22 antocianidinas são conhecidas (FRANCIS, 2000), dentre elas, apenas seis estão presentes em alimentos: pelargonidina, cianidina, delphinidina, peonidina, petunidina e malvidina, que se distinguem entre si pelo número de hidroxilas e pelo grau de metoxilação no anel B, conforme apresentado na Figura 4 (SCALBERT; WILLIAMSON, 2000; LIMA; GUERRA, 2003).

Dentre as frutas vermelhas, como o morango e amora, o mirtilo apresenta maior capacidade antioxidante, a qual é diretamente ligada ao seu alto teor de antocianinas (KALT *et al.*, 1999). Lohachoompol e colaboradores (2008) identificaram e quantificaram antocianinas em várias cultivares de mirtilo produzidas na Austrália. As cultivares estudadas foram: Crunchie, Star e Sharpe (“*highbush*”, *Vaccinium corymbosum*); Clímax, Powderblue e Brightwell (“*rabbiteye*”, *Vaccinium ashei*). Foram identificadas as antocianinas: delphinidina-3-glicosídeo, delphinidina-3-galactosídeo, cianidina-3-galactosídeo, delphinidina-3-arabinosídeo, cianidina-3-glicosídeo, petunidina-3-galactosídeo, cianidina-3-arabinosídeo, petunidina-3-glicosídeo, peonidina-3-galactosídeo, petunidina-3-arabinosídeo, peonidina-3-glicosídeo, malvidina-3-galactosídeo, peonidina-3-arabinosídeo, malvidina-3-glicosídeo e malvidina-3-arabinosídeo. As antocianidinas majoritárias encontradas foram a delphinidina, petunidina e malvidina.

#### 2.4.5 Atividades biológicas

Estudos epidemiológicos indicam que o consumo de alimentos ricos em componentes bioativos está associado à redução de DCV, acidente vascular cerebral e câncer (BAGCHI *et al.*, 2004; PAPANDREOU *et al.*, 2009). O mirtilo é uma importante fonte alimentar de antocianinas, polifenóis e flavonóides e parecem ter a maior capacidade antioxidante entre frutas e legumes (PRIOR *et al.*, 2000), conferindo ao mirtilo o título de AF (CHO *et al.*, 2004; HUANG *et al.*, 2012).

Muitos estudos demonstram que os frutos mirtilo têm uma ampla gama de benefícios à saúde, tais como atividade antioxidante, tanto *in vitro* (LI *et al.*, 2013; CASTREJÓN, *et al.*, 2008) quanto *in vivo* (DULEBOHN *et al.*, 2008; MOLAN, 2008),

que pode ser associado ao seu alto teor de antocianinas (RASEIRA & ANTUNES, 2004). Atividades anti-hipertensiva (SHAUGHNESSY *et al.*, 2009; KALEA *et al.*, 2009), antiobesidade (PRIOR *et al.*, 2009), antidiabética (DEFURIA *et al.*, 2009; STULL *et al.*, 2010) e atividade antitumoral (YI, *et al.*, 2005; NETO, 2007) também foram descritas. DeFuria *et al.*, (2009) mostraram em seu estudo que a expressão de genes inflamatórios foram reduzidos em ratos após o consumo de mirtilo, sugerindo uma resposta anti-inflamatória.

As folhas de mirtilo também tem atraído a atenção de pesquisadores uma vez que tem sido relatado que os polifenóis (especialmente proantocianidinas) podem suprimir a expressão do vírus da hepatite C (TAKESHITA, *et al.*, 2009). As folhas de mirtilo tem demonstrado exercer também atividade antimicrobiana e antioxidante, pois possuem uma grande quantidade de compostos bioativos, como os flavonóides (LI *et al.*, 2013).

Inoue e colaboradores, em 2011, analisaram o efeito da infusão das folhas de mirtilo sobre o perfil lipídico e o acúmulo de triglicerídeos hepáticos em ratos com sobrepeso. O extrato das folhas, obtidas a partir de uma infusão, demonstraram um efeito hipolipemiante e os autores atribuem o efeito as proantocianidinas e flavonóides presentes nas folhas.



### 3.0 OBJETIVOS

#### 3.1 Objetivo geral:

Investigar vias de estresse oxidativo em ratos Wistar hipercolesterolêmicos suplementados com o extrato dos frutos de *Vaccinium ashei* Reade.

#### 3.2 Objetivos específicos:

- Obter o extrato liofilizado dos frutos de *Vaccinium ashei* Reade;
- Avaliar o potencial antioxidante do extrato de mirtilo *in vitro*, através da determinação da atividade scavenger do radical DPPH e do radical ABTS;
- Quantificar o conteúdo de polifenóis totais do extrato liofilizado;
- Traçar o perfil lipídico (colesterol total e frações e triglicerídeos), glicêmico, hematológico e marcadores cardíacos, antes e após 14 dias de suplementação;
- Determinar a atividade das enzimas antioxidantes em eritrócitos: catalase (CAT), superóxido dismutase (SOD) e glutathione peroxidase (GPx);
- Obter os níveis de vitamina C em plasma;
- Determinar o conteúdo de polifenóis em plasma;
- Determinar o dano oxidativo em proteínas plasmáticas;
- Avaliar o nível de peroxidação lipídica através da medida das espécies reativas ao ácido tiobarbitúrico (TBARS) em plasma;
- Determinar o dano oxidativo no DNA de leucócitos do sangue periférico, através do ensaio cometa e micronúcleo;
- Análise de cortes histológicos de aortas.

## **PARTE II**

### **MANUSCRITO I**

Em fase de preparação para submissão para The Journal of Nutrition

#### **Hypolipidemic Effect of *Vaccinium ashei* Reade (blueberry) in Experimentally Induced Hypercholesterolemic Wistar Rats**

Deise Jaqueline Ströher, Ritiele Pinto Coelho, Angélica Aparecida da Costa Güllich,  
Bruna Cocco Pilar, Jamila Benvegnú Bruno, Jacqueline da Costa Escobar Piccoli,  
Vanusa Manfredini.

## **Hypolipidemic Effect of *Vaccinium ashei* Reade (blueberry) in Experimentally Induced Hypercholesterolemic Wistar Rats**

Deise Jaqueline Ströher<sup>a</sup>, Ritiele Pinto Coelho<sup>b</sup>, Angélica Aparecida da Costa Güllich<sup>a</sup>, Bruna Cocco Pilar<sup>a</sup>, Jamila Benvegnú Bruno<sup>b</sup>, Jacqueline da Costa Escobar Piccoli<sup>c</sup>, Vanusa Manfredini<sup>a,b,\*</sup>.

<sup>a</sup> Postgraduate Program in Biochemistry, Federal University of Pampa, Uruguaiiana, Rio Grande do Sul, Brazil.

<sup>b</sup> Course of Pharmacy, Federal University of Pampa, Uruguaiiana, Rio Grande do Sul, Brazil.

<sup>c</sup> Postgraduate Program in Pharmaceutical Sciences, Federal University of Pampa, Uruguaiiana, Rio Grande do Sul, Brazil.

RUNNING TITLE: Hypolipidemic Effect of *Vaccinium ashei* Reade in Hypercholesterolemic Rats

WORD COUNT: 3.460; NUMBER OF FIGURES: 2; NUMBER OF TABLES: 3

SUPPLEMENTARY MATERIAL: Online Supporting Materials: 0

AUTHOR LIST FOR INDEXING: Ströher, Coelho, Güllich, Pilar, Bruno, Piccoli, Manfredini.

\*To Whom Correspondence should be addressed, e-mail:

vanusamanfredini@unipampa.edu.br

Current address:

<sup>a,b</sup> Universidade Federal do Pampa – Campus Uruguaiana, Laboratório de Hematologia e Citologia Clínica, BR 472, Km 585, Uruguaiana, RS, Brazil, CEP: 97500-970. Tel.: 5555 3413-4321. Fax: 55553414-1484.

<sup>c</sup> Universidade Federal do Pampa – Campus Uruguaiana, Laboratório de Histologia, BR 472, Km 585, Uruguaiana, RS, Brazil, CEP: 97500-970. Tel.: 5555 3413-4321. Fax: 55553414-1484.

**ABSTRACT**

Blueberry (*Vaccinium ashei* Reade) is a fruit rich in bioactive compounds such as polyphenols, especially anthocyanins, and have different pharmacological properties. In the present study, we found that supplementation for 2 weeks with lyophilized extract of blueberry (BE), decreased the level of total cholesterol, LDL cholesterol and triglycerides, as well as increased plasma levels of HDL cholesterol in hypercholesterolemic rats. When the extract was associated to the simvastatin, the hypocholesterolemic effect demonstrated to be better, especially on the group receiving blueberry extract at a dose of 50 mg/Kg associated with simvastatin. Moreover, we observed that treatment with BE, protected against weight gain. However, no significant change was observed in the glycemic and hematology profile. The BE also showed an antiatherogenic effect, which may be promising for the development of new drugs.

*Keywords:* blueberry, hypercholesterolemia, lipid profile, atherosclerosis.

## 1.0 INTRODUCTION

Hypercholesterolemia has been implicated in atherosclerosis, which is the leading cause of death among world populations. Atherosclerosis is considered as a chronic and progressive disease, arising from the inflammatory processes and oxidative stress within vessel wall.<sup>1,2</sup> High cholesterol diet increases serum low-density lipoprotein (LDL) levels results in increased oxidized LDL levels.<sup>3</sup> Oxidation of LDL acting as a strong pro-atherogenic factor by triggering a complex inflammatory process<sup>4</sup> that triggers accumulation of macrophage white blood cells in the artery wall. Rupture of the plaque deposits oxidized cholesterol into the artery wall leading to atherosclerotic plaque formation.<sup>5,6</sup>

The 3-hydroxymethylglutaryl coenzyme A reductase (HMG CoA reductase) inhibitors, or statins, are a widely used group of hypocholesterolemic drugs, which are effective in reducing atherosclerotic cardiovascular events, largely by reducing plasma LDL concentrations.<sup>7</sup> The Simvastatin is one of the most commonly prescribed statins worldwide.<sup>8</sup>

Dietary flavonoids have emerged as potential candidates to protect against cardiovascular disease (CVD).<sup>9</sup> Epidemiological studies associate regular consumption of flavonoid-rich foods and beverages with a decreased risk of CVD mortality, which is mainly due to the potential of these bioactive components in to increasing serum antioxidant capacity and thereby protect against LDL oxidation and prevent CVD.<sup>10</sup>

There has been a growing interest in natural products as an alternative to pharmaceutical medications and their contribution to maintenance or improvement of health. The cholesterol-lowering effects of dietary plants has been well studied and various plants were shown to be helpful in lowering plasma cholesterol levels<sup>11,12</sup> and are considered to be useful means to prevent disorders such as atherosclerosis.<sup>13</sup>

Blueberries are an important dietary source of anthocyanins, polyphenols and flavonoids and appear to have the highest antioxidant capacity among fruits and vegetables,<sup>14</sup> conferring on blueberries the title of a functional food<sup>15,16</sup> and creating an opportunity for their use in the nutraceutical industry.<sup>17</sup>

The blueberry (*Vaccinium* spp) is a fruit species native to parts of Europe and North America where the fruit is considered as "source of longevity".<sup>18</sup> Several studies provide evidence of antioxidative,<sup>19</sup> antiinflammatory,<sup>20,21</sup> antihypertensive,<sup>22,23</sup> antiobesity,<sup>24</sup> and, antidiabetic<sup>25,26</sup> effects of blueberries.

Considering the important role of elevated plasma levels of lipids in the development of insulin resistance, atherosclerosis and cardiovascular disease<sup>27</sup> and there are however no reports on the effect of *Vaccinium ashei* Reade on hypercholesterolemia, this work was therefore aimed to investigate the effect of extract of fruits of *Vaccinium ashei* R on biochemical, hematological and histopathological profile in hypercholesterolemic rats.

## **2.0 MATERIALS AND METHODS**

### **2.1 Sample plant**

Mature fruits of rabbiteye blueberry (*Vaccinium ashei* Reade) cultivars Bluegem were collected in november of the year 2012 and transported fresh to the city of Uruguaiana, RS where they were immediately processed. The fruits were generously provided by EMBRAPA – Clima Temperado, Pelotas, Brazil, RS, established through collaboration with Maria do Carmo Bassols Raseira.

### **2.2 Preparation of the extract**

For the preparation of the extract were used 100g of blueberry macerated using a porcelain grail and pistil, in the dark, to preserve the antioxidant properties of its constituents

and then mixed with 100 mL of methanol: ethanol: acetone (45:45:10 v/v) according to the method described by Vizzotto and Pereira (2009)<sup>28</sup>, with adaptations. After 24 hours the extract was filtered and subjected to reduced pressure in a rotary evaporator to remove the solvent. The extract was transferred to Falcon tubes, frozen and subsequently lyophilized. The lyophilized extract was stored at -70°C until treatment.

### **2.3 Animal Experimentation**

For this study we used male Wistar rats (60-65 g), 30 days old, were obtained from the Central Animal Laboratory of the Federal University of Santa Maria, Rio Grande do Sul, Brazil. During treatment, rats were housed at a constant room temperature, humidity, and light cycle (12:12h light-dark), free access to tap water and fed with standard chow *ad libitum*. All experiments were conducted in compliance with the guidelines for biomedical research stated by the Brazilian Societies of Experimental Biology and approved by the Ethics Committee on Animal Use Experimentation of the Federal University of Pampa, CEUA, Urugaiana, Rio Grande do Sul, Brazil (Protocol n ° 035/2012).

### **2.4 Preparation of feed and Induction of hypercholesterolemia**

Commercial feed was supplemented with 7,5% of pig grease and with 2,5 % corn oil for each 100 g of feed according to the described methodology of Fietz and Salgado (1999)<sup>29</sup>, with adaptations. The hyperlipidemic feed was prepared daily for all experimental groups.

After the induction of the hypercholesterolemia, the groups kept receiving the same diet until the end of the experiment. 44 rats that consumed the hypercholesterolemic diet were selected. From them, 2 rats were euthanized after the first week of hypercholesterolemia induction to be used as comparison parameter, and were dosed the blood levels of total cholesterol, LDL-cholesterol, HDL-cholesterol and triglycerides. This procedure was



developed weekly until the hypercholesterolemia confirmation, totalizing 30 days of induction.

## **2.5 Experimental Desing**

The 36 rats were divided into 6 groups of 6 rats each. The group 1 (control) received saline solution in a daily dose of 1 mL, group 2: oral suspension of simvastatin 10 mg/Kg (SIM), group 3: blueberry extract 25 mg/Kg (BE25), group 4: blueberry extract 50 mg/Kg (BE50), group 5: blueberry extract 25 mg/Kg associated to simvastatin 10 mg/Kg (BE25+SIM), group 6: blueberry extract 50 mg/Kg associated to simvastatin 10 mg/Kg (BE50+SIM). All treatments were administered by gavage. The register of the body weight of the animals was performed during all experimental period to accompany the development of the animals and to determine the extracts and administrated drugs volume.

## **2.6 Blueberry extract prepare and administration**

The blueberry-based solution was obtained by the dissolution of the extract previously lyophilized, in water. The solution was daily prepared, in the moment immediately before the administration. The extracts were administered by gavage for 14 consecutive days. Animals were euthanized 24 h after the last treatment, in fasting, to obtain the whole blood and aorta.

## **2.7 Biochemical, hematological and cardiac markers analysis**

The hemograms (complete blood count) were performed in an automatic counter Cell-Dyn 3200 Hematology Analyzer (Abbott Diagnostic, St Clara, CA, USA) and total cholesterol, HDL-cholesterol, triglycerides total, glucose and creatine-kinase (CK) levels using automatic analyzers A-25 Biosystems (Biosystems SA, Barcelona, Spain) for *in vitro* diagnostics. Creatine kinase-MB (CK-MB) using Architect Abbott for *in vitro* diagnostics. Determination of plasma total homocysteine using HPLC coupled to mass spectrometry (LC-MS/MS), according to the technique described by Nelson *et al.*, (2003)<sup>30</sup>. LDL cholesterol

values were computed according to the Friedewald formula. All biochemical assays were carried out in triplicate.

## **2.8 Collection of Aorta and Histopathological analysis**

After 14 days of treatment, the animals were euthanized and the thoracic abdominal cavity was opened. The heart together with the aorta (2–3 cm length) was excised from each animal. The aorta was cut at the origin and removed from the heart. The entire aortas were rapidly dissected out and tissue sections (5 mm) fixed by immersion at room temperature in 10% formalin solution. For the histological examinations, paraffin embedded tissue sections of aorta were stained with hematoxylin-eosin (H&E). The tissue samples were for examined and photographed under a light microscope for observation of structural abnormality. The aortic diameter measurements were made with the program Image Pro-Plus.

## **2.9 Statistical Analysis**

Data were expressed as mean  $\pm$  standard deviation (SD). Comparisons between groups were performed using a two-way analysis of variance (ANOVA), followed by post hoc of Bonferroni for multiple comparison tests. Results were considered statistically significant when  $p < 0.05$ . The statistical analysis was performed using the software GraphPad Prism version 5 (GraphPad Software, Inc., La Jolla, CA, USA).

## **3.0 RESULTS**

### **3.1 Effects of BE on body weight gain**

During the period of hypercholesterolemia induction (week 1–4) there has not been significant difference on the body weight gain among rats. After 4 weeks, the hypercholesterolemic rats received the treatments along with the hypercholesterolemic diet. At the end of 2 weeks, the treatment of the group which received simvastatin (SIM) showed a significant decrease ( $p < 0.05$ ) of the body weight when compared to the control group. The

groups that received the blueberry extract (BE25 and BE50) demonstrated a significant decrease of the body weight ( $p < 0.05$ ) when compared to the control groups and the SIM. The groups which received the BE associated to the simvastatin (BE25+SIM e BE50+SIM) showed a significant decrease ( $p < 0.05$ ) of the body weight when compared to the group which received SIM (**Table 1**).

### 3.2 Effect of the BE treatment on Biochemical Profile

The effect of BE administration, for 2 weeks on lipid and glycemie profile in rats with induced hypercholesterolemia is illustrated in **Figure 1**. The BE25, BE50 and BE25+SIM administration along with the hypercholesterolemic diet was able to significantly decrease ( $p < 0.05$ ) serum total cholesterol levels compared with that of control and SIM groups. The BE50+SIM treated group also showed a significantly decrease ( $p < 0.05$ ) in serum total cholesterol levels with that of control group, but the treated group BE50+SIM was comparable to the SIM in terms of cholesterol lowering effect (**Figure 1A**).

The HDL-cholesterol levels significantly increased ( $p < 0.05$ ) in the group treated with BE dose of 50 mg/Kg (BE50) and the BE25+SIM group when compared to the control group, but the group treated with BE50+SIM showed an increase in HDL-cholesterol less when compared BE50 and BE25+SIM groups (**Figure 1B**).

The BE50+SIM administration showed significantly decrease ( $p < 0.05$ ) LDL-cholesterol levels with compared that of control, SIM and BE25, BE50 groups, showing a better effect in lowering LDL-cholesterol than the SIM alone (**Figure 1C**).

The triglycerides levels significantly decreased ( $p < 0.05$ ) in all treated groups compared with the control group, but the groups treated with the BE or SIM showed no significant difference between them (**Figure 1D**).

Treatment with BE showed no significant differences among the groups in glycaemic profile (**Figure 1E**).

### **3.3 Effect of the BE treatment on Hematological Profile**

The plasma platelets counts were higher in hypercholesterolemic rats. However, the results showed significant decrease in plasma levels of platelets and leukocytes in the group treated with BE25+SIM. The neutrophils also decreased in the group treated with BE50+SIM. The other parameters evaluated showed no significant results (**Table 2**).

### **3.4 Effect of the BE treatment on Markers of cardiac damage**

The creatine-kinase (CK) and its isoform CK-MB are used as cardiac disease marker. The hypercholesterolemic diet induced an increase on the CK-total levels on the control group, although at the end of 14 days of treatment, the group which received SIM showed a decrease ( $p<0.05$ ) on the CK-total levels when compared to the control group. The groups that received BE25 and BE50 also showed a decrease ( $p<0.05$ ) on the CK-total levels when compared to the control group and SIM. The CK-total levels also demonstrated a statistically significant decrease ( $p<0.05$ ) on the groups which received the BE25+SIM and BE50+SIM demonstrating a better result than the groups that received only the extract. The group that received SIM has not showed significant differences on the CK-MB levels when compared to the control group. The groups that received BE with doses of 25 mg/Kg and 50 mg/Kg and the groups which received the same doses associated to the simvastatin showed a statistically significant decrease ( $p<0.05$ ) on the CK-MB levels when compared to the control groups and SIM. The hypercholesterolemic diet also induced to a significant increase ( $p<0.05$ ) on the serum levels of homocysteine on the control group when comparing to the groups treated with the BE or SIM. The groups that received SIM, BE25 and BE25+SIM showed a statistically significant decrease ( $p<0.05$ ) on the levels of homocysteine, compared to the group which

only received a hypercholesterolemic diet (control group). The groups which received the isolated extract of 50 mg/Kg (BE50) and the group that homocysteine received the extract of 50 mg/Kg associated to (BE50+SIM) demonstrated a statistically significant decrease similar to the serum levels of homocysteine, when compared to the other groups. The results are showed on **Table 3**.

### **3.5 Effect of the BE treatment on Histopathological Profile**

Histopathological analysis of the aorta of hypercholesterolemic rats showed spaces of fat droplets within the tunica intima and media. These findings indicate that hypercholesterolemia disturbed the prooxidant-antioxidant balance in favor of peroxidation in the aorta tissues together with atherosclerotic changes in the aorta of rats. However, histopathological observation showed significant decrease of aortic lesions in the BE25, BE50, BE25+SIM and BE50+SIM groups when compared to control and SIM groups, and the improvement is more evident when the extract (50mg/Kg) is associated with simvastatin. **(Figure 2)**

## **4.0 DISCUSSION**

Hypercholesterolemia is a great concern in the occidental countries as the main etiology for the atherosclerosis and cardiovascular diseases (CVD). The consumption of a rich diet on cholesterol increases the level of lipid peroxidation, which is also one of the initial processes of atherosclerosis. Studies have demonstrated that some antioxidants are associated to an anti-hypercholesterolemic effect and can prevent the atherosclerosis, protecting the LDL from oxidation.<sup>31,32</sup>

The statins are a category of drugs widely used to decrease cholesterol and it has been proved to be efficient on the reduction of morbidity and mortality of these two conditions related to high levels of cholesterol. Additional effects of the statins have been demonstrated

and showed to be important on the stabilization of the atherosclerotic plaque. These effects (also called pleiotropic), refer to the endothelial protection, to the reduction of the lipid peroxidation and to the control of the inflammatory and hemostasis.<sup>33,34,35,36</sup> However, other studies have demonstrated the security profile of the statins and some concerns came up about the collateral effects that this category of drugs may provoke in a long term.<sup>37</sup>

Our results showed that the groups which received treatment with blueberry extract in doses of 25 mg/Kg and 50 mg/kg (BE25 and BE50) for 2 weeks showed a significantly decrease in body weight. This result corroborates the findings of Song *et al.*, 2013<sup>38</sup>, we used a HFD-induced obesity rat model to investigate the anti-obesity effects of blueberry extract. The groups receiving blueberry extract at the dosage of 60 mg/kg and 150 mg/kg orally for 5 weeks and the result showed that the body weight of rats were significantly lower.

Studies show that the natural antioxidants, as the polyphenols, can be used for the effective correction of high levels of total cholesterol and triglycerides in the blood.<sup>39</sup> In our study the groups treated with blueberry extract were able to significantly reduce levels of total cholesterol and triglycerides, when compared to the control group. Again, these data support the study of Song *et al.*, 2013<sup>38</sup> indicating that the blueberry extract effectively regulates the metabolism of cholesterol and triglycerides in obese rats induced by hypercholesterolemic diet.

Coffy, in 2008<sup>40</sup>, carried out a study with hypercholesterolemic rats treated with blueberry extract, showed serum reduction on LDL-cholesterol levels on the percentage of 46% and the increase of the HDL-cholesterol in about 10%. On this study, the blueberry extract on the doses of 25 mg/Kg e 50 mg/Kg (BE25 e BE50), it was capable of reducing the total cholesterol and the LDL-cholesterol in a significant way. When the extract was

associated to the simvastatin, the hypercholesterolemic effect demonstrated to be better, specially on the group BE50+SIM.

In our study, the blueberry extract (BE) was also capable of diminishing the level of serum triglycerides of the hypercholesterolemic rats and proved to have an activity similar to the isolated simvastatin. Inoue and collaborators, in 2011<sup>41</sup>, analyzed the effect of the infusion of the blueberry leaves on the lipid profile and the accumulation of hepatic triglycerides on overweight rats. The leaves extract, obtained from an infusion, showed an hypolipemic effect and the authors attach this effect to the presence of the proanthocyanidins and flavonoids present on the leaves.

The HDL-cholesterol performs an important role on the defense against oxidative damage of the membranes.<sup>42,43</sup> The main role of the HDL on the lipid metabolism is the absorption and cholesterol transportation from the peripheral tissues to the liver through a reverse transportation of cholesterol. Diminished levels of HDL-cholesterol are strongly associated to a high risk of CDV.<sup>44,45</sup> In our study, the HDL-cholesterol levels increased in the groups that received the treatment with blueberry extract and it was significantly ( $p < 0.05$ ) higher in the group that received the extract on the dose of 50 mg/Kg. These results prove the favorable performance of the BE on the lipid profile of *Wistar* rats.

However, in our study, the BE has not showed effect on the fasting glycaemia, datum also found by Basu and collaborators in 2010.<sup>46</sup>

The supplementation with fruits, vegetables and seeds that contain flavonoids, contribute to the hypolipidemic effect. The blueberry is an important feeding source of antocianins, polyphenols and flavonoids<sup>47</sup>, and these bioactive compounds are probably the responsible for the reduction of the hypercholesterolemia.

The CK has been used as a cardiac disease marker. The increase of the induced cholesterol by the diet has increased the levels of the CK-total and CK-MB on the control group, although in the groups that received treatment with BE, there have been a significant decrease of the enzyme levels. However, the group that received the BE50+SIM showed a meaningful improvement when compared to the isolated extract. The high levels of homocysteine in the blood are also associated to an increased risk of CVD.<sup>48</sup> In this case, there have been significant differences of the groups that received the BE from the ones which have not received. Based on the results achieved, we can state that the blueberry had an important role in the prevention of cardiac damage of the rats submitted to the hypercholesterolemic diet.

Histopathological analysis of the aorta of hypercholesterolemic rats showed significant decrease of aortic lesions in groups treated with BE. These results suggest a protective effectiveness of blueberry extract against atherosclerosis in hypercholesterolemic rats. The potential mechanisms may involve reduction in oxidative stress by both inhibition of lipid peroxidation and an enhancement of antioxidant defense. Wu *et al.*, 2010<sup>49</sup> have reported that dietary supplementation with 1% freeze-dried blueberries (BB) for 20 weeks decreased aortic lesions. The authors accordingly have suggested that the atheroprotective effect of BB may have been related to its antioxidative effect.

In conclusion, the BB supplementation in hypercholesterolemic rats showed protect against weight gain, improve the lipid profile, reversing the high levels of total cholesterol, LDL and triglycerides, and showed an antiatherogenic effect. These results show that blueberry have positive effects, and therefore, they may have potential for use in the development of functional food or nutraceuticals.



## **ACKNOWLEDGEMENTS**

This work was supported by grants from UNIPAMPA (Universidade Federal do Pampa) UFRGS (Universidade Federal do Rio Grande do Sul), FAPERGS (Fundação de Amparo a Pesquisa do Estado do Rio Grande do Sul), CAPES (Coordenação de Aperfeiçoamento de Pessoal de Nível Superior) and CNPq (Conselho Nacional de Desenvolvimento Científico e Tecnológico).

## 5.0 LITERATURE CITED

1. HANSSON, G.K. Inflammation, atherosclerosis, and coronary artery disease. *N Engl J Med* **2005**, 1685-1695.
2. DORIA, A.; SHERER, Y.; MERONI, P.L.; SHOENFELD, Y. Inflammation and accelerated atherosclerosis: basic mechanisms. *Rheum Dis Clin North Am*, **2005**; *31*,355-62.
3. WARNHOLTZ, A.; MOLLNAU, H.; OELZE, M.; WENDT, M.; MUNZEL, T. Antioxidants and endothelial dysfunction in hyperlipidemia. *Current Hypertension Reports*, **2001**, *3*, 53-60.
4. YOUNIS, N.; CHARLTON-MENYS, V.; SHARMA, R.; SORAN, H.; DURRINGTON, P.N. Glycation of LDL in non-diabetic people: small dense LDL is preferentially glycosylated both in vivo and in vitro. *Atherosclerosis*, **2009**, *202*, 162–168.
5. AVIRAM, M. Review of human studies on oxidative damage and antioxidant protection related to cardiovascular diseases. *Free Radic. Res.* **2000**, *33*, S85–97.
6. AVIRAM, M.; KAPLAN, M.; ROSENBLAT, M.; FUHRMAN, B. Dietary antioxidants and paraoxonases against LDL oxidation and atherosclerosis development. *Handb. Exp. Pharmacol*, **2005**, *170*, 263–300.
7. BAIGENT, C.; BLACKWELL, L.; EMBERSON, J. *et al.* Efficacy and safety of more intensive lowering of LDL cholesterol: a meta-analysis of data from 170,000 participants in 26 randomised trials. *Lancet*, **2010**,1670–1681.
8. HU, MIAO.; VALIANT, W.L.; MARK, YAJIE.; XIAO & BRIAN TOMLINSON. Associations between the genotypes and phenotype of CYP3A and the lipid response to simvastatin in Chinese patients with hypercholesterolemia. *Pharmacogenomics*, **2013**, *1*, 25–34.

9. WALLACE, T.C. Anthocyanins in cardiovascular disease. *Adv. Nutr.* **2011**, 2, 1–7.
10. HE, J.; & GIUSTI, M. M. Anthocyanins: Natural Colorants with Health-Promoting Properties. *Food Science and Technology*, **2010**, 1, 163-187.
11. KIM, B.; KU C.S.; PHAM, T.X.; PARK, Y.; MARTIN, D.A.; XIE, L.; TAHERI, R.; LEE, J.; BOLLING, B.W. Aronia melanocarpa (chokeberry) polyphenol-rich extract improves antioxidant function and reduces total plasma cholesterol in apolipoprotein E knockout mice. *Nutr Res.* **2013**, 33, 406-13.
12. LIANG, Y.; CHEN, J.; ZUO, Y.; MA, K. Y.; JIANG, Y.; HUANG, Y.; CHEN, Z. Y. Blueberry anthocyanins at doses of 0.5 and 1 % lowered plasma cholesterol by increasing fecal excretion of acidic and neutral sterols in hamsters fed a cholesterol-enriched diet. *European journal of nutrition*, **2013**, 52, 869–875.
13. HAKIMOGLU, FIDAN; KIZIL, GOKSEL; KANAY, ZEKI; KIZIL, MURAT; ISI, HILMI. The effect of ethanol extract of *Hypericum lysimachioides* on lipid profile in hypercholesterolemic rabbits and its *in vitro* antioxidant activity. *Atherosclerosis*, **2007**, 192, 113-122.
14. PRIOR, R.L.; CAO, G.; PRIOR, R.L.; CAO, G. Analysis of botanicals and dietary supplements for antioxidant capacity: a review. *JAOAC Int*, **2000**, 83, 950–956.
15. CHO, M. J. et al. Flavonoid glycosides and antioxidant capacity of various blackberry, blueberry and red grape genotypes determined by high-performance liquid chromatography/mass spectrometry. *Journal of the Science of Food and Agriculture*, **2004**, v. 84, n. 13, p. 1771-1782.
16. HUANG, W. Y. et al. Survey of antioxidant capacity and phenolic composition of blueberry, blackberry, and strawberry in Nanjing. *Journal of Zhejiang University Science B*, **2012**, v. 13, n. 2, p. 94-102.

17. LI, C.; FENG, J.; HUANG, W.Y.; AN, X.T. Composition of polyphenols and antioxidant activity of rabbiteye blueberry (*Vaccinium ashei*) in Nanjing. *J Agric Food Chem*, **2013**, *23*, 523–531.
18. RASEIRA, M. B. C.; ANTUNES, L.E.C. **A Cultura do mirtilo**. Pelotas: Embrapa Clima Temperado, 2004.
19. BRAGA, P.C.; ANTONACCI, R.; WANG, Y.Y.; LATTUADA, N.; DAL SASSO, MARABINI, M. L.; FIBIANI, M.; LO SCALZO, R. Comparative antioxidant activity of cultivated and wild *Vaccinium* species investigated by EPR, human neutrophil burst and COMET assay. *Eur Rev Med Pharmacol Sci*, **2013**; *17*, 1987-1999.
20. AHMET, I.; SPANGLER, E.; SHUKITT-HALE, B.; JUHASZOVA, M.; SOLLOTT, S.J.; JOSEPH, J.A.; INGRAM, D.K.; TALAN, M. Blueberry-enriched diet protects rat heart from ischemic damage. *PLoS ONE*. **2009**; *4*, 5954.
21. LAU, F.C.; BIELINSKI, D.F.; JOSEPH, J.A. Inhibitory effects of blueberry extract on the production of inflammatory mediators in lipopolysaccharide activated BV2 microglia. *J Neurosci Res*. **2007**, *85*,1010–7.
22. SHAUGHNESSY, K.S.; BOSWALL, I.A.; SCANLAN, A.P.; GOTTSCHALL-PASS, K.T.; SWEENEY, M.I. Diets containing blueberry extract lower blood pressure in spontaneously hypertensive stroke-prone rats. *Nutr Res*. **2009**; *29*, 130–8.
23. KALEA, A.Z.; CLARK, K.; SCHUSCHKE, D.A.; KLIMIS-ZACAS, D.J. Vascular reactivity is affected by dietary consumption of wild blueberries in the Sprague-Dawley rat. *J Med Food*. **2009**;12:21.
24. PRIOR, R.L.; WU, X.; GU, L.; HAGER, T.; HAGER, A.; WILKES, S.; HOWARD, L. Purified berry anthocyanins but not whole berries normalize lipid parameters in mice fed an obesogenic high fat diet. *Mol Nutr Food Res*. **2009**; *53*, 1406–18.

25. DEFURIA, J.; BENNETT, G.; STRISSEL, K.J.; PERFIELD, J.W.I.I.; MILBURY, P.E.; GREENBERG, A.S.; OBIN, M.S. Dietary blueberry attenuates whole-body insulin resistance in high fat-fed mice by reducing adipocyte death and its inflammatory sequelae. *J Nutr.* **2009**; *139*,1510–6.
26. STULL, A. J.; CASH, K. C.; JOHNSON, D. J.; CHAMPAGNE, C. M.; CEFALU, W. T. Bioactive in blueberries improve insulin sensitivity in obese, insulin-resistant men and women. *J. Nutr.* **2010**, *140*, 1764-1768.
27. BARBALHO, S. M. et al. *Mentha piperita* effects on wistar rats plasma lipids. *Brazilian Archives of Biology and Technology*, **2009**, *52*: 1137-1143.
28. VIZZOTTO, M.; Metodologia científica: otimização do processo de extração de compostos fenólicos antioxidantes de mirtilo (*Vaccinium ashei* Reade). *Boletim de Pesquisa e Desenvolvimento*. Embrapa Clima Temperado, Pelotas, **2009**. URL ([http://www.cpact.embrapa.br/publicacoes/download/boletins/boletim\\_101.pdf](http://www.cpact.embrapa.br/publicacoes/download/boletins/boletim_101.pdf)).
29. FIETZ, V.R.; SALGADO, J.M. Efeito da pectina e da celulose nos níveis séricos de colesterol e triglicérides em ratos hiperlipidêmicos. *Ciênc. Tecnol. Aliment.* **1999**, *19*, 3.
30. NELSON, B.; PFEIFFER, C.; SNIEGOSKY, L.; SATTERFIELD, M. Development and evaluation of an isotope dilution LC/MS method for the determination of total homocysteine in human plasma. *Analytical Chemistry*. **2003**, v.75, n.4, p.775-784.
31. CHEN, M.F.; HSU, H.C.; LIAU, C.S.; LEE, Y.T. The role of Vitamin E on the antiatherosclerotic effect of fish oil in diet-induced hypercholesterolemic rabbits. *Prostaglandins other Lipid Mediators*, **1999**; *57*:99–111.
32. FREYSCHUSS, A.; AL-SCHURBAJI, A.; BJÖRKHEM, I.; et al. On the antiatherogenic effect of the antioxidant BHT in cholesterol-fed rabbits: inverse relation between serum triglycerides and atheromatous lesions. *Biochim Biophys Acta*, **2001**; *1534*:129–38.

33. RICKER, P.M.; RIFAIL, N.; LOWENTHAL, S.P. Rapid reduction in C-reactive protein with cerivastatin among 785 patients with primary hypercholesterolemia. *Circulation*, **2001**; 103:1191-3.
34. BELLOSTA, S.; BERNINI, F.; FERRI, N.; QUARATO, P.; CANAVESI, M.; ARNABOLDI, L. Direct vascular effects of HMGCoA reductase inhibitors. *Atherosclerosis*, **1998**.137:S101-S109.
35. SCALIA, R.; APPEL, J.Z.; LEFER, A.M. Leukocyte-endothelium interaction during the early stages of hypercholesterolemia in the rabbit. *Arterioscl Thromb Vasc Biol*, **1998**, 18,1093-100.
36. DANGAS, G.J.; BADIMON, L.; SMITH, D.A.; LEVINE, A.; AMBROSE, J.A. Pravastatin therapy in hyperlipidemia. Effects on thrombus formation and the systemic hemostatic profile. *Am Coll Cardiol*, **1999**; 33:1294-304.
37. MCKENNEY, J.M.; DAVIDSON, M.H.; JACOBSON, T.A.; GUYTON, J.R. Final conclusions and recommendations of the National Lipid Association Statin Safety Assessment Task Force. *Am J Cardiol*, **2006**, 97, 89C–94C.
38. SONG, Y.; PARK, H.J.; KANG, S.N.; JANG, S.H.; LEE, S.J.; KO, Y.G.; KIM, G.S.; CHO, J.H. Blueberry peel extracts inhibit adipogenesis in 3T3-L1 cells and reduce high-fat diet-induced obesity. *PLoS One*, **2013**, 8, 7.
39. CHANANDIRI, T.; SANIKIDZE, T.; ESAISHVILI, M.; CHKHIKVISLI, I.; DATUNASHVILI, I. Effectiveness of green tea catechines for the correction of the alimentary obesity in the experiment. Inst of Medical Biotechn, *Georgian Academy of Scien*. **2005**, 61 n. 3 p. 126.
40. COFFY, S.P. Nível sérico de NO e TBARS em ratos Wistar submetidos à dieta hipercolesterolêmica tratados com extratos de diferentes espécies de *Vaccinium myrtillus*.

**2008.** 25f. Trabalho de conclusão de curso (Graduação em Farmácia) – Universidade regional Integrada do Alto Uruguai e das Missões, Erechim.

41. INOUE, N., NAGAO, K.; NOMURA, S.; SHIROUCHI, B.; INAFUKU, M.; HIRABARU, H.; NAKAHARA, N.; NISHIZONO, S.; TANAKA, T.; YANAGITA, T. Effect of *Vaccinium ashei* reade leaf extracts on lipid metabolism in obese OLETF rats. *Biosci Biotechnol Biochem*, **2011**, *75*, 2304-2308.

42. NOFER, J.R.; KEHREL, B.; FOBKER, M.; LEVKAU, B.; ASSMANN, G.; VON ECKARDSTEIN, A. HDL and arteriosclerosis: beyond reverse cholesterol transport. *Atherosclerosis* **2002**, *161*,1–16.

43. PARTHASARATHY, S.; BARNETT, J.; FONG, L.G. High density lipoprotein inhibits the oxidative modification of lowdensity lipoprotein. *Biochim Biophys Acta*, **1990**;1044:275–83.

44. FARIAS, R.A.F.; NETO, M.F.O.; VIANA, G.S.B.; RAO, V.S.N.I. Effects of *Croton cajucara* extract on serum lipids of rats fed a high fat diet. *Phytother Res*, **1996**, *10*,697–9.

45. WILSON, P.W.; ABBOTT, R.D.; CASTELLI, W.P. High-density lipoprotein cholesterol and mortality. The Framingham Heart study. *Atherosclerosis*, **1988**, *8*, 737–41.

46. BASU, A.; SANCHEZ, K.; LEYVA, M. et al.: Green tea supplementation affects body weight, lipids, and lipid peroxidation in obese subjects with metabolic syndrome. *J Am Coll Nutr* **2010**; *29*:31–40.

47. PRIOR, R.L.; LAZARUS, S.A.; CAO, G.; MUCCITELLI, H.; HAMMERSTONE, J.F. Identification of procyanidins and anthocyanins in blueberries and cranberries (*Vaccinium* spp.) using high-performance liquid chromatography/mass spectrometry. *J. Agric. Food Chem*, **2001**, *49*, 1270–1276.

48. MARTI-CARVAJAL, A.J.; SOLA, I.; LATHYRIS, D.; SALANTI, G. Homocysteine lowering interventions for preventing cardiovascular events. *Cochrane Database Syst Rev*, **2009**, CD006612.
49. WU, X. et al. Dietary blueberries attenuate atherosclerosis in apolipoprotein E-deficient mice by upregulating antioxidant enzyme expression. *Journal of Nutrition*, **2010**, *140*, 1628-1632.



## FIGURE LEGENDS

**Figure 1:** Effect of blueberry extract in hypercholesterolemic rats after treatment. In A: serum total cholesterol levels; B: HDL-cholesterol levels; C: LDL-cholesterol levels; D: triglycerides levels; E: glucose levels. Data are expressed as means±S.D. Different letters are significantly ( $p<0.05$ ) different by two-way ANOVA followed by Bonferroni's comparison pos hoc test.

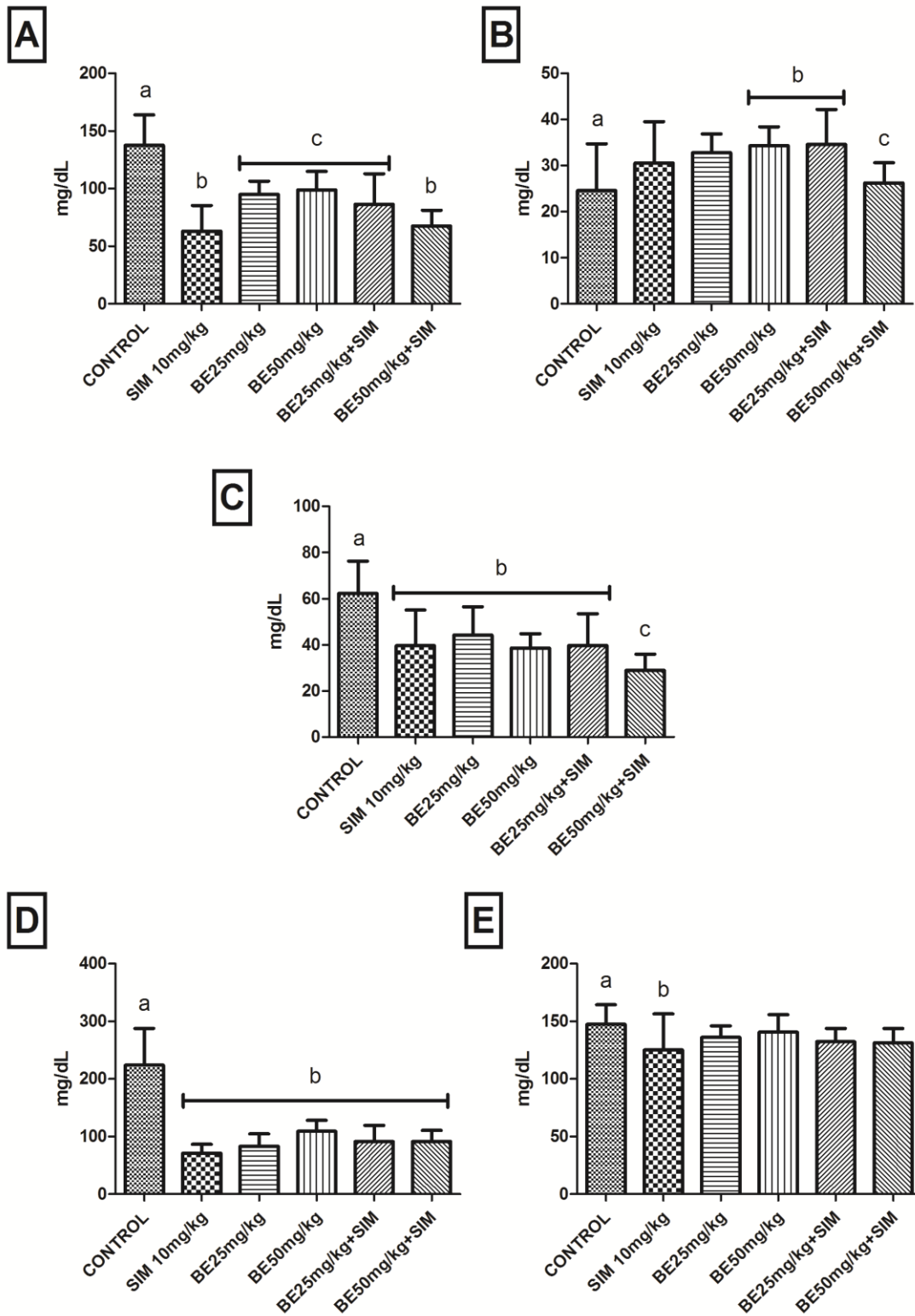
**Figure 2:** Histological section and thickening measure of the aorta of hypercholesterolemic rats after treatment. In 1: control group (100x); 1B: simvastatin (200x); 1C: blueberry extract 25 mg/Kg (100x); 1D: blueberry extract 50 mg/Kg (40x); 1E: blueberry extract 25 mg/Kg + simvastatina 10 mg/Kg (40x); 1F: blueberry extract 50 mg/Kg + simvastatina 10 mg/Kg (40x). (H&E). In 2: Measure aortic thickening ( $\mu\text{m}$ ) in groups subjected to different treatments. Different letters are significantly different by two-way ANOVA followed by Bonferroni's comparison pos hoc test. Different letters are significantly ( $p<0.05$ ) different by two-way ANOVA followed by Bonferroni's comparison pos hoc test.

**Table 1** - Body weight of rats during the induction of hypercholesterolemia (weeks 1-4) and during treatment (weeks 5-6).

GROUPS	BODY WEIGHT					
	INDUCTION OF HYPERCHOLESTEROLEMIA				TREATMENT	
	Week 1	Week 2	Week 3	Week 4	Week 5	Week 6
<b>CONTROL</b>	131.0 ± 3.79	159.9 ± 9.00	180.1 ± 11.71	210.2 ± 13.19	238.3 ± 12.72	260.3 ± 12.80 <sup>a</sup>
<b>SIM</b>	127.9 ± 4.41	148.7 ± 24.98	159.0 ± 24.02	191.1 ± 28.04	183.1 ± 55.47	189.3 ± 70.34 <sup>b</sup>
<b>BE 25</b>	130.5 ± 3.07	158.2 ± 6.12	174.4 ± 10.38	208.2 ± 10.56	235.7 ± 15.95	253.6 ± 20.17 <sup>b,c</sup>
<b>BE 50</b>	131.5 ± 4.08	160.1 ± 10.43	179.0 ± 13.45	213.4 ± 18.19	242.2 ± 16.43	257.3 ± 16.17 <sup>b,c</sup>
<b>BE 25+SIM</b>	128.8 ± 3.79	158.2 ± 9.08	169.0 ± 11.43	204.9 ± 6.25	224.8 ± 10.02	242.6 ± 11.06 <sup>c</sup>
<b>BE 50+SIM</b>	129.5 ± 5.49	159.0 ± 14.59	168.6 ± 13.10	200.8 ± 14.59	217.9 ± 12.33	233.9 ± 18.92 <sup>c</sup>

Values are expressed as Mean ± SD of each group (n=6). Different letters are significantly different by two-way ANOVA followed by Bonferroni's comparison pos hoc test.

Figure 1



**Table 2** – Hematological parameters of the hypercholesterolemic rats exposed to different treatments.

PARAMETERS	GROUPS					
	CONTROL	SIM	BE 25	BE 50	BE 25+SIM	BE 50+SIM
Hemoglobin (g/dL)	13.58 ± 2.71	13.82±1.54	13.90±0.53	12.90±3.31	12.96±2.98	15.15±0.90
Hematocrit (%)	39.83±9.17	40.56±4.27	40.60±1.33	41.10±1.82	42.83±2.64	43.35±2.19
MCV (fL)	55.17±1.72	54.40±0.82	55.67±1.13	57.33±1.53	56.20±1.65	54.75±1.12
MHC (pg)	19.48±1.09	18.56±0.42	19.05±0.51	20.12±0.66	19.24±0.87	19.20±0.50
MCHC (%)	35.28±1.91	34.10±0.40	34.18±0.33	35.23±0.96	19.24±0.87	19.20±0.50
Erythrocytes (10 <sup>6</sup> /mm <sup>3</sup> )	6.49±2.08	7.45±0.81	7.29±0.23	7.17±0.39	6.80±1.74	7.76±0.27
Leukocytes (10 <sup>3</sup> /mm <sup>3</sup> )	5.76±1.23	2.60±0.95 <sup>a</sup>	6,58±2,25	6,98±1,93	3,84±1,87 <sup>b</sup>	6,30±0,53
Neutrophils (%)	22,17±3,43	25,00±7,12	23,17±7,11	21,40±6,05	24,60±7,98	14,50±3,68 <sup>a</sup>
Lymphocytes (%)	72,83±2,17	69,75±7,17	73,50±7,43	69,17±11,93	70,40±6,56	79,25±4,15
Eosinophils (%)	2,33±0,76	2,20±0,41	1,66±0,76	1,66±0,76	2,20±0,77	2,50±0,89
Monocytes (%)	1,83±0,70	2,60±0,50	1,66±0,76 <sup>a</sup>	3,50±0,78	3,40±1,40	2,75±0,44
Basophil (%)	0.00±0.00	0.00±0.00	0.16±0.38	0.16±0.38	0.00±0.00	0.00±0.00
Platelets (10 <sup>3</sup> /mm <sup>3</sup> )	730,2±246,8	741,5±211,9	814,4±114,5	713,7±132,5	379,7±332,3 <sup>a</sup>	786,0±92,97

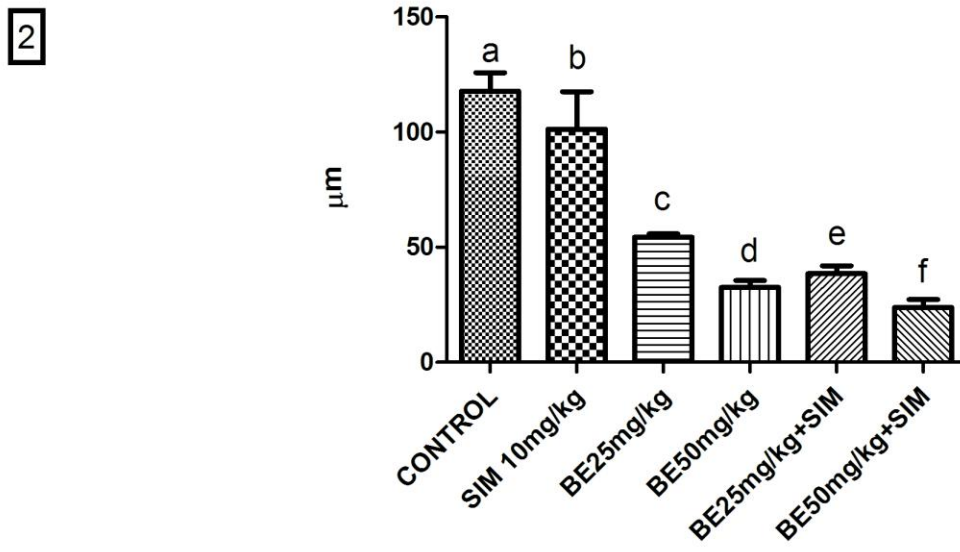
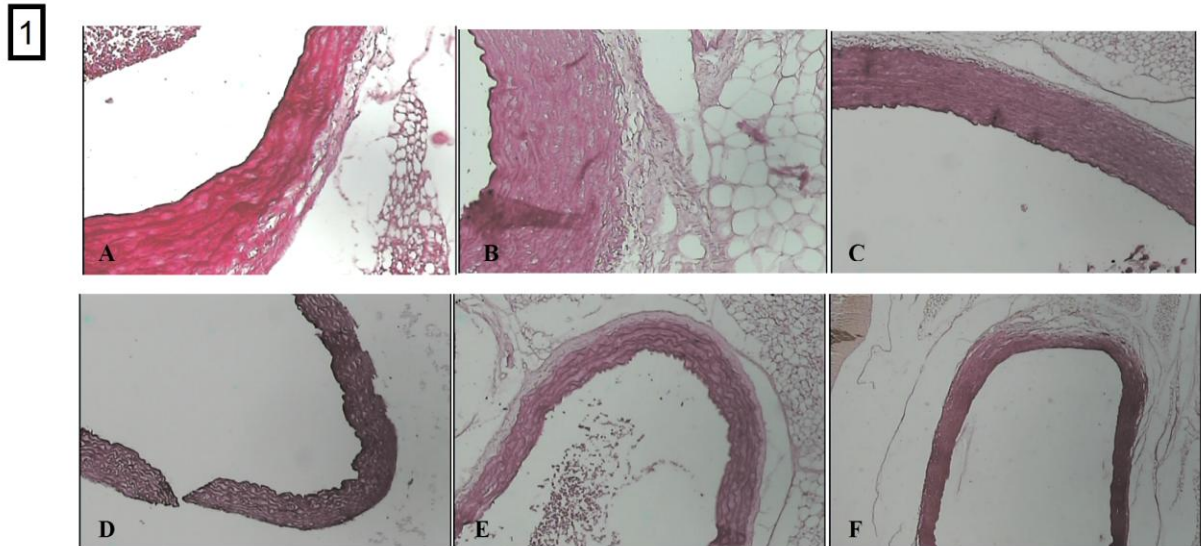
Data are expressed as means±SD. Different letters are significantly ( $p<0.05$ ) different by two-way ANOVA followed by Bonferroni's comparison pos hoc test.

**Table 3** – Markers of cardiac damage in the hypercholesterolemic rats exposed to different treatments.

<b>GROUPS</b>	<b>CK-TOTAL (U/L)</b>	<b>CK-MB (U/L)</b>	<b>HOMOCYSTEINE (<math>\mu</math>mol/L)</b>
<b>CONTROL</b>	132.5 $\pm$ 39.67 <sup>a</sup>	4.72 $\pm$ 0.80 <sup>a</sup>	17.52 $\pm$ 0.48 <sup>a</sup>
<b>SIM</b>	83.60 $\pm$ 7.30 <sup>b</sup>	4.36 $\pm$ 0.35 <sup>a</sup>	11.02 $\pm$ 0.22 <sup>b</sup>
<b>BE 25</b>	66.33 $\pm$ 2.11 <sup>c</sup>	3.16 $\pm$ 0.09 <sup>b</sup>	10.77 $\pm$ 0.11 <sup>b</sup>
<b>BE 50</b>	49.67 $\pm$ 5.84 <sup>d</sup>	2.35 $\pm$ 0.18 <sup>c</sup>	8.98 $\pm$ 0.06 <sup>c</sup>
<b>BE 25 + SIM</b>	27.40 $\pm$ 1.54 <sup>e</sup>	1.58 $\pm$ 0.18 <sup>d</sup>	10.80 $\pm$ 0.06 <sup>b</sup>
<b>BE 50 + SIM</b>	21.67 $\pm$ 5.46 <sup>e</sup>	0.90 $\pm$ 0.21 <sup>e</sup>	8.49 $\pm$ 0.04 <sup>c</sup>

Means with different letters at a time differ significantly,  $p < 0.05$ . The values sharing common letters are not significantly different at  $p < 0.05$ . CK-total: serum total creatine kinase. CK-MB: CK isoform. Different letters are significantly different by two-way ANOVA followed by Bonferroni's comparison pos hoc test.

Figure 2



**MANUSCRITO II**

Em fase de preparação para submissão para Journal of Agricultural and Food Chemistry

**Antioxidant effect of *Vaccinium ashei* Reade (blueberry) extract in hypercholesterolemic rats**

Deise Jaqueline Ströher, Ritiele Pinto Coelho, Angélica Aparecida da Costa Güllich, Bruna Cocco Pilar, Jamila Benvegnú Bruno, Jacqueline da Costa Escobar Piccoli, Vanusa Manfredini.

**Antioxidant effect of *Vaccinium ashei* Reade (blueberry) extract in hypercholesterolemic rats**

Deise Jaqueline Ströher<sup>a</sup>, Ritiele Pinto Coelho<sup>b</sup>, Angélica Aparecida da Costa Güllich<sup>a</sup>, Bruna Cocco Pilar<sup>a</sup>, Jamila Benvegnú Bruno<sup>b</sup>, Jacqueline da Costa Escobar Piccoli<sup>c</sup>, Vanusa Manfredini<sup>a,b,\*</sup>.

<sup>a</sup> Postgraduate Program in Biochemistry, Federal University of Pampa, Uruguaiana, Rio Grande do Sul, Brazil.

<sup>b</sup> Course of Pharmacy, Federal University of Pampa, Uruguaiana, Rio Grande do Sul, Brazil.

<sup>c</sup> Postgraduate Program in Pharmaceutical Sciences, Federal University of Pampa, Uruguaiana, Rio Grande do Sul, Brazil.

\*To Whom Correspondence should be addressed: Vanusa Manfredini, Universidade Federal do Pampa – Campus Uruguaiana, Laboratório de Hematologia e Citologia Clínica, BR 472, Km 585, Uruguaiana, RS, Brazil, CEP: 97500-970. Tel.: 5555 3413-4321. Fax: 55553414-1484

E-mail: vanusamanfredini@unipampa.edu.br



## ABSTRACT

Blueberries are an important dietary source of polyphenols, anthocyanins, flavonoids and has greater antioxidant capacity than most other fruits and vegetables, being considered a functional food. *Vaccinium ashei* lyophilized extract was administered in diet-induced hypercholesterolemic rats during 2 weeks and parameters of oxidative stress, antioxidant enzymes, polyphenols and vitamin C were measured. The current study demonstrated that blueberry extract (BE) has antioxidant potential *in vitro* as verified by DPPH radical and ABTS scavenging activities as well as by the high total polyphenols content. In addition, treatment with the BE showed a decrease of lipid peroxidation and protein and an increase in activities of antioxidant enzymes like catalase (CAT), superoxide dismutase (SOD) and glutathione peroxidase (GPx). A significant increase in the levels of vitamin C and polyphenols in plasma also were found. Dietary consumption of polyphenols rich foods may contribute to overall antioxidant status, particularly in reducing oxidative stress associated with hypercholesterolemia.

*Keywords:* *Vaccinium ashei* Reade, hypercholesterolemia, polyphenols, oxidative stress.

## 1.0 INTRODUCTION

Oxidative stress has been demonstrated to play a causal role in different vascular diseases, such as hypertension, diabetic, hypercholesterolemia and atherosclerosis<sup>1</sup>. Reactive oxygen species (ROS), such as superoxide, hydrogen peroxide, and hydroxyl radicals, and free radical-mediated reactions, can cause oxidative damage to cellular structures and functional molecules (e.g., DNA, proteins, and lipids), contributing to the development of cardiovascular diseases<sup>2</sup>. Dietary compounds such as polyphenols, found in fruits and vegetables, can play an important role in the improvement of antioxidant status because they are able to neutralize ROS<sup>3,4</sup>.

With the current upsurge of interest in the efficacy and use of naturally derived antioxidants, functional foods have received much attention in recent years<sup>5</sup>. Blueberries are known as “super fruits” for their potential in the nutraceutical markets<sup>6,7</sup> because contain anthocyanins, polyphenols and flavonoids beyond a high level of vitamin C (ascorbic acid), folic acid and resveratrol<sup>8</sup> and appear to have the highest antioxidant capacity among fruits and vegetables, conferring on blueberries the title of a functional food<sup>9,5</sup>.

Many reports have suggested that blueberry fruits have a wide range of health benefits such as antioxidant activity *in vivo*<sup>10</sup> and *in vitro*<sup>11</sup>, antidiabetic activity<sup>12,13</sup>, and the ability to protect against cancer and stroke<sup>14</sup>.

With the current upsurge of interest in the efficacy and use of naturally derived antioxidants, functional foods and nutraceuticals in recent years, and, considering that Brazil has recently become a blueberry producer with a small production<sup>15</sup> the objectives of this study were to evaluate the role of blueberry (*Vaccinium ashei* R) on oxidative stress parameters and antioxidant defenses in hypercholesterolemic rats after treatment of blueberry extract.

## **2.0 MATERIALS AND METHODS**

### **2.1 Chemicals**

All the chemicals were from Sigma Chemical Co. (St. Louis, MO, USA) and of analytical grade.

### **2.2 Plant Material**

Mature fruits of rabbiteye blueberry (*Vaccinium ashei* Reade) cultivars Bluegem were collected in november of the year 2012 and transported fresh to the city of Uruguaiana, RS where they were immediately processed. The fruits were generously provided by EMBRAPA – Clima Temperado, Pelotas, Brazil, RS, established through collaboration with Maria do Carmo Bassols Raseira.

### **2.3 Preparation of the extract**

For the preparation of the extract were used 100 g of blueberry macerated using a porcelain grail and pistil, in the dark, to preserve the antioxidant properties of its constituents and then mixed with 100 mL of methanol: ethanol: acetone (45:45:10 v/v) according to the method described by Vizzotto and Pereira (2009)<sup>16</sup>, with adaptations. After 24 hours the extract was filtered and subjected to reduced pressure in a rotary evaporator to remove the solvent. The extract was transferred to Falcon tubes, frozen and subsequently lyophilized. The lyophilized extract was stored at -70°C until treatment.

### **2.4 Evaluation of antioxidant potential from blueberry extract in vitro**

In order to determine the antioxidant potential of this obtained blueberry extract we evaluated the scavenging activity of 2,2-diphenyl-1-picrylhydrazyl (DPPH) radical, 2,2'-azino-bis-3-ethylbenzthiazoline-6- sulphonic acid (ABTS) as well as we quantified the total polyphenols content.

#### 2.4.1. DPPH radical scavenging activity

The DPPH (2,2-diphenyl-1-picrylhydrazyl) radical-scavenging activity of blueberry extract was determined as described by Sharma and Bhat<sup>17</sup>. The DPPH radical solution (50 $\mu$ L) was dissolved in methanol and added to a medium containing blueberry extract at different concentrations (0.5-10,1 mg/mL). The medium was incubated at room temperature for 30 min. The decrease in absorbance was measured at 517 nm, which depicted the scavenging activity of blueberry extract against DPPH. The DPPH scavenging capacity of the compound was calculated as

$$\text{DPPH radical scavenging activity} = 100 - [(ABS_{\text{SAMPLE}} - ABS_{\text{BLANK}}) / ABS_{\text{CONTROL}}] \times 100$$

Where,  $ABS_{\text{SAMPLE}}$  is the absorbance of the test compound,  $ABS_{\text{BLANK}}$  is the absorbance of the blank and  $ABS_{\text{CONTROL}}$  is the absorbance of the control reaction.  $IC_{50}$  value (concentration of sample where absorbance of ABTS decreases 50% with respect to absorbance of blank) of the sample was determined. Ascorbic acid was used as positive control.

#### 2.4.2 ABTS radical scavenging assay:

The ABTS method (2,2'-azino-bis-3-ethylbenzthiazoline-6-sulphonic acid) is based on the deactivation of the antioxidant radical cation  $ABTS^{\bullet+}$ , which is measured by the decrease in absorbance at 734 nm. The ABTS method was performed as described by Re *et al.*<sup>18</sup>. Absorbance was read at 734 nm, 7 minutes after the extract addition. The ABTS scavenging capacity of the compound was calculated as

$$\text{ABTS radical scavenging activity} = 100 - [(ABS_{\text{SAMPLE}} - ABS_{\text{BLANK}}) / ABS_{\text{CONTROL}}] \times 100$$

Where,  $ABS_{\text{SAMPLE}}$  is the absorbance of the test compound,  $ABS_{\text{BLANK}}$  is the absorbance of the blank and  $ABS_{\text{CONTROL}}$  is the absorbance of the control reaction.  $IC_{50}$  value (concentration of sample where absorbance of ABTS decreases 50% with respect to absorbance of blank) of the sample was determined. Ascorbic acid was used as positive control.

### **2.4.3 Determination of total polyphenols content**

Total polyphenols content of the blueberry extract was measured by spectrophotometry using the Folin-Ciocalteu method<sup>19</sup>, with modifications. Briefly, 1 mL of 1 N Folin-Ciocalteu reagent was added to a 1 mL of sample, and this mixture was allowed to stand for 2-5 min before the addition of 2 mL of 20% Na<sub>2</sub>CO<sub>3</sub>. The solution was then allowed to stand for 10 minutes before reading at 750 nm in Spectrophotometer (UV-1800 Shimadzu, Japan) using 1 cm quartz cells. The total polyphenol content was expressed as milligram of gallic acid equivalent per milliliter (mg GAE mL<sup>-1</sup>).

## **2.5 Evaluation of blueberry extract in vivo**

### **2.5.1 Animals Experimentation**

For this study we used male Wistar rats (60-65 g), 30 days old, were obtained from the Central Animal Laboratory of the Federal University of Santa Maria, Rio Grande do Sul, Brazil. During treatment, rats were housed at a constant room temperature, humidity, and light cycle (12:12h light-dark), free access to tap water and fed with standard chow *ad libitum*. All experiments were conducted in compliance with the guidelines for biomedical research stated by the Brazilian Societies of Experimental Biology and approved by the Ethics Committee on Animal Use Experimentation of the Federal University of Pampa, CEUA, Uruguaiana, Rio Grande do Sul, Brazil (Protocol n° 035/2012).

### **2.5.2 Preparation of feed and Induction of hypercholesterolemia**

Commercial feed was supplemented with 7,5% of pig grease and with 2,5 % corn oil for each 100g of feed according to the described methodology of Fietz and Salgado (1999), with adaptations. The hyperlipidemic feed was prepared daily for all experimental groups.

After the induction of the hypercholesterolemia, the groups kept receiving the same diet until the end of the experiment. 44 rats that consumed the hypercholesterolemic diet were

selected. From them, 2 rats were euthanized after the first week of hypercholesterolemia induction to be used as comparison parameter, and were dosed the blood levels of total cholesterol, LDL-cholesterol, HDL-cholesterol and triglycerides. This procedure was developed weekly until the hypercholesterolemia confirmation, totalizing 30 days of induction.

### **2.5.3 Experimental Desing**

The 36 rats were divided into 6 groups of 6 rats each. The group 1 (control) received saline solution in a daily dose of 1 mL, group 2: oral suspension of simvastatin 10 mg/Kg (SIM), group 3: blueberry extract 25 mg/Kg (BE25), group 4: blueberry extract 50 mg/Kg (BE50), group 5: blueberry extract 25 mg/Kg associated to simvastatin 10 mg/Kg (BE25+SIM), group 6: blueberry extract 50 mg/Kg associated to simvastatin 10 mg/Kg (BE50+SIM). All treatments were administered by gavage. The register of the body weight of the animals was performed during all experimental period to accompany the development of the animals and to determine the extracts and administrated drugs volume.

### **2.5.4 Blueberry extract prepare and administration**

The blueberry-based solution was obtained by the dissolution of the extract previously lyophilized, in water. The solution was daily prepared, in the moment immediately before the administration. The extracts were administered by gavage for 14 consecutive days. Animals were euthanized 24 h after the last treatment, in fasting, to obtain the whole blood.

## **2.6 Evaluation of oxidative stress parameters**

### **2.6.1 Oxidative damage**

Oxidative damage markers, lipid peroxidation<sup>21</sup>, protein carbonyls<sup>22</sup> in plasma were measured by the spectrophotometric methods. The assessment of DNA damage was made by comet assay<sup>23</sup> and frequency of micronucleus<sup>24</sup> in leukocytes.

### 2.6.2 Antioxidant defenses

The activities of superoxide dismutase (SOD), catalase (CAT) and glutathione peroxidase (GPx) in erythrocyte were achieved using commercial kits (Randox Brazil LTDA).

### 2.7 Statistical Analysis

Data were expressed as mean  $\pm$  standard deviation (SD). Comparisons between groups were performed using a two-way analysis of variance (ANOVA), followed by post hoc of Bonferroni for multiple comparison tests. Results were considered statistically significant when  $p < 0.05$ . The statistical analysis was performed using the software GraphPad Prism version 5 (GraphPad Software, Inc., La Jolla, CA, USA).

## 3.0 RESULTS/DISCUSSION

### 3.1 Radical scavenging capacity of lyophilized blueberry extract

Antioxidant activity of the tested lyophilized blueberry extract (*Vaccinium ashei* R) was determined by both DPPH and ABTS methods and showed good antioxidant capacity. The results are shown in **Figure 1**.

We verified that the lyophilized blueberry extract (0.5 and 10 mg/mL) demonstrated a significant DPPH scavenging activity (2.5 and 52.43 % of inhibition, respectively) (**Figure 1A**) and significant ABTS scavenging activity (4.8 and 73.78 % of inhibition, respectively) (**Figure 1B**). The obtained data illustrated that, ABTS method showed higher antioxidant activity than DPPH method, because the ABTS method, which is more sensitive than the DPPH method.

The lyophilized blueberry extract analyzed in this study showed highest antioxidant activity: DPPH,  $IC_{50}$  9.57 mg/mL and ABTS,  $IC_{50}$  6.85 mg/mL. These results confirm the blueberry extract as a source of phenol compounds with high antioxidant activity.

### 3.2 The total polyphenols content

We detected that the polyphenols content in the lyophilized blueberry extract was 190.620 mg GAE/mL.

### 4.1 Evaluation of oxidative stress parameters

#### 4.1.2 Oxidative damage

The incidence of atherosclerosis increases with hypercholesterolemia.<sup>25</sup> Oxidative modifications in proteins, lipids, and DNA are considered to be among the molecular mechanisms leading to endothelial dysfunction and atherosclerosis.<sup>26</sup>

The **Figure 2** shows the results of biomarkers for oxidative damage. The malondialdehyde (MDA) has the potential not only to assess the extent of oxidative damage, but also to predict potential efficacy of therapeutic strategies aimed at reducing oxidative stress.<sup>27</sup>

Hypercholesterolemia is associated with elevated levels of MDA and is also known to increase the production of ROS. In this study plasma MDA level was used to investigate the effect of the blueberry extract on hypercholesterolemic rats. MDA level was measured as TBARS (thiobarbituric acid reactive substances) method. This method has been criticized for its lack of specificity, but it is one of the easiest and the most frequently used marker of lipid peroxidation.<sup>28</sup>

Several investigators found that high cholesterol diet had an increasing effect on lipid peroxidation in plasma and tissues.<sup>29,30</sup> The results of this study show significant increases ( $p<0.05$ ) in plasma MDA levels in control group when compared with other groups. However, the groups treated with BE25 and BE50+SIM show significant decreases ( $p<0.05$ ) MDA levels in plasma when compared with control group (**Figure 2A**). This result suggests that this BE may have an antioxidant effect on hypercholesterolemic rats because the polyphenols,



present in BE, are incorporated into membrane lipid and act as hydrogen donors, trapping free radicals and inhibiting the formation of lipid radicals.<sup>31</sup> The antioxidant effects of blueberries have also been reported by studies using cellular and animal models of oxidative stress.<sup>32,33</sup>

Protein carbonylation is a type of protein oxidation that can be promoted by ROS. It usually refers to a process that forms reactive ketones or aldehydes that can be reacted by 2,4-dinitrophenylhydrazine (DNPH) to form hydrazones. Direct oxidation of side chains of lysine, arginine, proline, and threonine residues, among other amino acids, in the “primary protein carbonylation” reaction produces DNPH detectable protein products.<sup>34</sup> To assess the effects of BE supplementation on hypercholesterolemic rats, the levels of carbonyl proteins measured and the results shown in **Figure 2B**. Protein carbonyls are normally used as a biomarker for protein damage caused by oxidized amino acid residues in stress conditions.<sup>35</sup>

We observed a significant effect of the dietary intervention with BE in protect the proteins against the reactive species. This effect is enhanced when the extract is associated with simvastatin (BE50+SIM). This result corroborated those showed by Aydin *et al.*, 2009<sup>36</sup>, comparison of cholesterol-fed rabbits for 8 weeks with rabbits fed cholesterol fed (8 weeks) plus atorvastatin administration (4 weeks) revealed that a significant decreases in protein carbonyl and MDA in atorvastatin-treated rabbits. Atorvastatin therapy caused significant decreases in both protein carbonyl levels.

In **Figure 2C** is showed the result of micronucleus frequency. Our data show no alteration in these parameter.

The ROS can damage DNA, lipids, and proteins, it is necessary to measure damage of more than just one of these cellular components. DNA damage is the most severe form of oxidative damage because it can cause permanent mutations that are passed on to progeny cells. The comet assay was performed to determine damage index DNA and the results were

shown in **Figure 2D**. We observed a potential effect of the BE in comet assay, a genotoxicity test which has been widely used in recent years to analyze protective effect on DNA damage. In our study the groups treated with BE showed a decrease DNA damage in leukocytes of rats hypercholesterolemics and this effect is enhanced when the extract is associated with simvastatin (BE50+SIM). This result corroborated those showed by Barros *et al.*<sup>37</sup>, showing DNA damage levels were significantly reduced in the hippocampal regions of mice that had been supplemented with a much lower level of 2.6-3.2 mg/kg of body weight of blueberry anthocyanins in drinking water for 30 days. Possible mechanisms for these genoprotective effects include protection of DNA from alkylation or formation of anthocyanin-DNA complexes, which stabilize the molecule against oxidative attack.<sup>38,39</sup>

#### **4.1.3 Antioxidant defenses**

In **Figure 3**, we show the results of antioxidant defenses biomarkers in plasma. Polyphenols are plant secondary metabolites, widely present in commonly consumed foods of plant origin, and they are accruing a body of evidence as bioactive components in a wide range of biological systems.<sup>40</sup> These compounds are considered to carry many potential beneficial health effects.

We observed a significant effect of the treatment with BE increase the polyphenol concentration in the groups BE 25, BE50 and BE50+SIM, showing that treatment with the extract was able to increase, after 14 days of treatment, the content of polyphenols in plasma of hypercholesterolemic rats (**Figure 3A**).

Ascorbic acid or vitamin C is one of the important water soluble vitamins, being associated to many health benefits. It acts as a cofactor in the enzymatic biosynthesis of hormones, being also a potent antioxidant, as it reduces nitrogen and ROS into stable molecules.<sup>41</sup>

After 14 days of treatment with the BE, there was a significant increase in the levels of ascorbic acid in plasma. This increase was more evident in the group that received the highest dose extract (50mg/kg), and the group that received the combined extract SIM. This result shows that the blueberry is a source of ascorbic acid and is able to increase the antioxidant capacity in hypercholesterolemic rats (**Figure 3B**).

To minimize the oxidative damage caused by ROS, cells possess a wide range of enzymatic systems including, glutathione peroxidase (GPx). The results of the antioxidant enzymes, superoxide dismutase (SOD), catalase (CAT) e glutathione peroxidase (GPx) in serum hypercholesterolemic rats is showed in **Figure 3**.

The activity of the antioxidant enzyme CAT is shown in **Figure 3C**. The groups that received SIM, BE25 and BE25+SIM showed increased serum activity of this enzyme. However, the dose of 50mg/kg showed an even better effect on the increase in CAT activity when compared to the control group.

SOD is the major cellular antioxidant defense enzyme against superoxides in vascular cells and the may act synergistically to combat the oxidative stress implicated in atherosclerosis.<sup>42</sup> In our study, after 14 days treatment with blueberry extract, the activity of this enzyme also increased, especially in the groups that received the dose of 50mg/Kg extract. Again, when the extract was associated with SIM, increased SOD activity was more significant (**Figure 3D**). Wu *et al.*, 2010<sup>43</sup>, in their study, also found the activity of SOD increased in female mice that received AIN-93G diet incorporated with 1% freeze-dried blueberry powder.

GPx is a selenoenzyme that plays a key role in protecting the organism from oxidative damage by catalyzing the reduction of harmful hydroperoxides with thiol cofactors.<sup>44</sup> In our study, the activity of GPx enzyme increased when compared to the control group, the groups

treated with SIM, BE25 and BE50+SIM. However, the groups that received a higher dose of extract (50mg/Kg), GPx activity had a greater antioxidant activity (**Figure 3E**).

Another interesting finding of this study was that the activity of antioxidant enzymes increased when the blueberry extract was associated with simvastatin.

The current study demonstrated a decrease of lipid peroxidation and protein as well as an increase of antioxidants enzymes, in hypercholesterolemic rats after 2 weeks of treatment with BE. These results are consistent with the hypothesis that flavonoids and other polyphenols can have effects in decrease oxidative damage and many studies have demonstrated high levels of these compounds in BE.<sup>45,46</sup>

The lyophilized extract of *Vaccinium ashei* R. was shown to have many benefits, which may be related high polyphenols levels, in particularly the anthocyanidins and proanthocyanidins, responsible for their very strong antioxidant activity.

In summary, hypercholesterolemia increases the oxidative stress in plasma. BE is efficient in slowing the progression of hypercholesterolemia induced oxidative stress and improving their functions but BE50+SIM is more effective than BE.

**ABBREVIATIONS USED**

**BE** – blueberry extract

**BE25** – blueberry extract 25mg/Kg

**BE50** – blueberry extract 50mg/Kg

**BE25+SIM** – blueberry extract 25mg/Kg associated with simvastatin

**BE50+SIM** - blueberry extract 50mg/Kg associated with simvastatin

**DPPH** - 2,2-diphenyl-1-picrylhydrazyl

**ABTS** - 2,2'-azino-bis-3-ethylbenzthiazoline-6-sulphonic acid

**CAT** - catalase

**SOD** – superoxide dismutase

**GPx** - glutathione peroxidase

**ROS** – reactive oxygen species

**DNA** - deoxyribonucleic acid

**SIM** - simvastatin

**MDA** - malondialdehyde

**TBARS** - thiobarbituric acid reactive substances

**DNPH** - 2,4-dinitrophenylhydrazine

## **ACKNOWLEDGMENT**

This work was supported by grants from UNIPAMPA (Universidade Federal do Pampa) UFRGS (Universidade Federal do Rio Grande do Sul), FAPERGS (Fundação de Amparo a Pesquisa do Estado do Rio Grande do Sul), CAPES (Coordenação de Aperfeiçoamento de Pessoal de Nível Superior) and CNPq (Conselho Nacional de Desenvolvimento Científico e Tecnológico).

#### 4.0 REFERENCES

1. Magenta, A., Greco, S., Gaetano, C., Martelli, F. Oxidative Stress and MicroRNAs in Vascular Diseases. *International Journal of Molecular Sciences*. **2013**, *14*, 17319-17346.
2. Finkel T, Holbrook N. J. Oxidants oxidative stress and the biology of ageing. *Nature*. **2000**, *408*, 239–47.
3. Lodovici, M.; Guglielmi, F.; Casalini, C.; Meoni, M.; Cheynier, V.; Dolaro, P. Antioxidant and radical scavenging properties in vitro of polyphenolic extracts from red wine. *Eur. J. Nutr.* **2001**, *40*: 74–77.
4. Manach, C.; Scalbert, A.; Morand, C.; Remesy, C.; Jimenez, L. Polyphenols: food sources and bioavailability. *Am. J. Clin. Nutr.* **2004**, *79*, 727–747.
5. Huang, W.Y., Zhang, H.C., Liu, W.X., Li, C.Y., 2012. Survey of antioxidant capacity and phenolic composition of blueberry, blackberry, and strawberry in Nanjing. *J. Zhejiang Univ.-Sci. B (Biomed. & Biotechnol.)*, **2013** (2): 94-102.
6. Ding, M.; Feng, R.; Wang, S.Y.; Bowman, L., Lu; Y., Qian, Y.; Castranova, V., Jiang, B.H., Shi, X. Cyanidin-3-glucoside, a natural product derived from blackberry, exhibits chemopreventive and chemotherapeutic activity. *J. Biol. Chem.*, **2006**, *25*:17359-17368.
7. Tulipani, S.; Mezzetti, B.; Capocasa, F.; Bompadre, S.; Beekwilder, J.; de Vos, C.H.R.; Capanoglu, E.; Bovy, A.; Battino, M. Antioxidants, phenolic compounds, and nutritional quality of different strawberry genotypes. *J. Agric. Food Chem.*, **2008**, *3*, 696-704.
8. Rimando, A.M.; Kalt, W.; Magee, J.B.; Dewey, J.; Ballington, J.R. Resveratrol, pterostilbene, and piceatannol in *Vaccinium* berries. *J. Agric. Food. Chem.* **2004**, *52*, 4713–9.
9. Cho, M.J.; Howard, L.R.; Prior, R.L.; Clark, J.R. Flavonoid glycosides and antioxidant capacity of various blackberry, blueberry and red grape genotypes determined by high-

performance liquid chromatography/mass spectrometry. *J. Sci. Food Agric.*, **2004**, *13*, 1771-1782.

10. Braga, P. C.; Antonacci, R.; Wang, Y. Y.; Lattuada, N.; Dal Sasso, M.; Marabini, L.; Fibiani, M.; Los Scalzo, R. *Comparative antioxidant activity of cultivated and wild Vaccinium species investigated by EPR, human neutrophil burst and COMET assay. Eur Rev Med Pharmacol Sci*, **2013**; *17*, 1987-1999.

11. Castrejón, A. D. R., Eichholz, I., Rohn, S., Kroh, L. W., Huyskens-Keil, S., Phenolic profile and antioxidant activity of highbush blueberry (*Vaccinium corymbosum* L.) during fruit maturation and ripening. *Food. Chem.*, **2008**, *109*, 564–572.

12. DeFuria, J.; Bennett, G.; Strissel, K.J.; Perfield, J.W.II.; Milbury, P.E.; Greenberg, A.S.; Obin, M.S. Dietary blueberry attenuates whole-body insulin resistance in high fat-fed mice by reducing adipocyte death and its inflammatory sequelae. *J. Nutr.* **2009**, *139*, 1510–6.

13. Stull, A. J.; Cash, K. C.; Johnson, D. J.; Champagne, C. M.; Cefalu, W. T. Bioactive in blueberries improve insulin sensitivity in obese, insulin-resistant men and women. *J. Nutr.*, **2010**, *140*, 1764-1768.

14. Wang, Y.; Chang, C. F.; Chou, J.; Chen, H. L.; Deng, X.; Harvey, B. K.; Cadet, J. L.; Bickford, P. C. Dietary supplementation with blueberries, spinach, or spirulina reduces ischemic brain damage. *Exp. Neurol.*, **2005**, *193*, 75–84.

15. Rodrigues, E.; Poerner, N.; Rockenbac, I. I.; Gonzaga, L. V.; Mendes, C. R.; Fett, R.. Phenolic compounds and antioxidant activity of blueberry cultivars grown in Brazil. *Ciência e Tecnologia de Alimentos*, **2011**, *31*, 911-917.

16. Vizzotto, M.; Metodologia científica: otimização do processo de extração de compostos fenólicos antioxidantes de mirtilo (*Vaccinium ashei* Reade). **Boletim de Pesquisa e**



**Desenvolvimento.** Embrapa Clima Temperado, Pelotas, 2009. URL ([http://www.cpact.embrapa.br/publicacoes/download/boletins/boletim\\_101.pdf](http://www.cpact.embrapa.br/publicacoes/download/boletins/boletim_101.pdf)).

17. Sharma, O.P.; Bhat, T.K. DPPH antioxidant assay revisited. *Food Chemistry*. **2009**, *113*, 1202-05.
18. Re, R.; Pellegrini, N.; Proteggente, A.; Pannala, A.; Yang, M.; Rice-Evans, C. Antioxidant activity applying an improved ABTS radical cation decolorization assay. *Free Radic. Biol. Med.* **1999**, *26*, 1231 – 1237.
19. Singleton, V. L.; Orthofer, R.; & Lamuela-Raventos, R. M. Analysis of total phenols and other oxidation substrates and antioxidants by means of Folin–Ciocalteu reagent. *Methods in Enzymology*, **1999**, *299*, 152–178.
20. Fietz, V.R.; Salgado, J.M. Efeito da pectina e da celulose nos níveis séricos de colesterol e triglicerídeos em ratos hiperlipidêmicos. *Ciênc. Tecnol. Aliment.* **1999**, *19*, 3.
21. Ohkawa, H.; Ohishi, N.; Yagi, K.; Assay for lipid peroxides in animal tissues by thiobarbituric acid reaction. *Anal. Biochem.* **1979**, *95*, 351-358.
22. Levine RL, Garland D, Oliver CN, Amici A, Climent I, Lenz AG, Alm B, Shaltiel S, Stadman ER. Damage to proteins and lipids tissues under oxidative stress. *Methods in Enzymology*. **1990**, *186*, 464-478.
23. Singh NP, McCoy MT, Tice RR and Schneider EL. A simple technique for quantitation of low levels of DNA damage in individual cells. *Experimental Cell Research*. **1988**, *175*, 184-91.
24. Schmid W. The micronucleus test. *Mutation Research*. **1975**, *31*: 9.
25. Çakatay, U.; Kayali, R.; Uzun, H. Relation of plasma protein oxidation parameters and paraoxonase activity in the ageing population. *Clin. Exp. Med.* **2008**, *8*, 51–7.

26. Aydin, S.; Uzun, H.; Sozer, V.; Altug, T. Effects of atorvastatin therapy on protein oxidation and oxidative DNA damage in hypercholesterolemic rabbits. *Pharmacol. Res.* **2009**, *59*, 242-247.
27. Yazdanparast, R.; Ardestani, A.; Jamshidi, S. Experimental diabetes treated with *Achillea santolina*: Effect on pancreatic oxidative parameters. *J. Ethnopharmacol.* **2007**, *112*, 13–18.
28. Yagi, K. Simple assay for the level of total lipid peroxides in serum or plasma. *Meth. Mol. Biol.* **1998**, *108*, 101–6.
29. Shukla, R.; Gupta, S.; Gambhir, J.K.; Prabhu, K.M.; Murthy, P.S.; Antioxidant effect of aqueous extract of the bark of *Ficus bengalensis* in hypercholesterolaemic rabbits. *J. Ethnopharmacol.* **2004**, *92*, 47–51.
30. Kim, B.J.; Kim, Y.K.; Park, W.H.; Ko, J.H.; Lee, Y.C.; Kim, C.H. A water-extract of the Korean traditional formulation Geiji–Bokryung–Hwan reduces atherosclerosis and hypercholesteremia in cholesterol-fed rabbits. *Int. Immunopharmacol.* **2003**, *3*, 723–34.
31. Soobrattee, M. A.; Neergheen, V. S.; Luximon-Ramma, A. e Aruoma, O. I., Bahorun, T. Phenolics as potential antioxidant therapeutic agents: Mechanism and actions. *Mutation Research*, **2005**, v. 579, p. 200-213.
32. Sellappan, S.; Akoh, C.C.; Krewer, G. Phenolic compounds and antioxidant capacity of Georgia-grown blueberries and blackberries. *J. Agric. Food. Chem.* **2002**, *50*, 432–8.
33. Ahmet, I.; Spangler, E.; Shukitt-Hale, B.; Juhaszova, M.; Sollott, S.J.; Joseph, J.A.; Ingram, D.K.; Talan, M. Blueberry-enriched diet protects rat heart from ischemic damage. *PLoS ONE*, **2009**, *4*, 5954.
34. Stadtman, E.R.; Levine, R.L. Protein oxidation. *Annals of the New York Academy of Sciences.* **2000**, *899*: 191-208.

35. Beal, MF. Oxidatively modified proteins in aging and disease. *Free. Radic. Biol. Med.* 2002, 32, 797–803.
36. Aydin, S.; Uzun, H.; Altug, T.; Effects of atorvastatin therapy on protein oxidation and oxidative DNA damage in hypercholesterolemic rabbits. *Clinical Biochemistry*, **2009**, 42, 4-5.
37. Barros, D.; Amaral, O. B.; Izquierdo, I.; Geracitano, L.; do Carmo Bassols Raseira, M.; Henriques, A. T.; Ramirez, M. R. Behavioral and genoprotective effects of *Vaccinium* berries intake in mice. *Pharmacol., Biochem. Behav.* **2006**, 84, 229–234.
38. Ramirez-Tortosa C, Andersen M, Gardner PT, Morrice PC, Wood SC, Duthie SJ, et al. Anthocyanin-rich extract decreases indices of lipid peroxidation and DNA damage in vitamin E-depleted rats. *Free Radic Biol Med* **2001**, 31 (9):1033–7.
39. Beattie J, Crozier A, Duthie GG. Potential health benefits of berries. *Curr. Nutr. Food Sci.* **2005**;1:71–86.
40. Ignat, I.; Volf, I.; Popa, V.I. A critical review of methods for characterisation of polyphenolic compounds in fruits and vegetables. *Food Chemistry.* **2011**, 126: 1821-35.
41. Wilson, J.X. The physiological role of dehydroascorbic acid. *FEBS Lett*, **2002**, 527, 5-9.
42. 't Hoen PA, Van der Lans CA, Van Eck M, Bijsterbosch MK, Van Berkel TJ, Twisk J. Aorta of ApoE-deficient mice responds to atherogenic stimuli by a prelesional increase and subsequent decrease in the expression of antioxidant enzymes. *Circ Res.* 2003;93:262–9.
43. Wu, X. et al. Dietary blueberries attenuate atherosclerosis in apolipoprotein E-deficient mice by upregulating antioxidant enzyme expression. *Journal of Nutrition*, v. **2010**, 140, p. 1628-1632.
44. Bhabak, K.P., Muges, G. Functional mimics of glutathione peroxidase: bioinspired synthetic antioxidants. *Acc Chem Res.* **2010**, 43, 1408-1419.

45. Ehlenfeldt, M. K.; Prior, R. L. Oxygen radical absorbance capacity (ORAC) and phenolic and anthocyanin concentrations in fruit and leaf tissues of highbush blueberry. *J. Agric. Food Chem.* **2001**, *49*, 2222–2227.
46. Su, M. S.; Silva, J. L. Antioxidant activity, anthocyanins, and phenolics of rabbiteye blueberry (*Vaccinium ashei*) by-products as affected by fermentation. *Food Chem.* **2006**, *97*, 447–451.

**FIGURE CAPTIONS**

**Figure 01:** Antioxidant activities *in vitro* of the lyophilized blueberry extract of *Vaccinium ashei* R determined at different concentrations: (A) DPPH radical-scavenging activity (B) ABTS radical-scavenging activity. AA – ascorbic acid; BE – blueberry extract.

**Figure 02:** Oxidative damage markers in hypercholesterolemic rats after treatment of blueberry extract. In A: lipid peroxidation levels; B: carbonyl protein contents; C: frequency of micronucleus; D: DNA damage index Data are expressed as means±S.D. Different letters means statistically different results ( $p<0.05$ ).

**Figure 03:** Antioxidative defenses markers in in hypercholesterolemic rats after treatment of blueberry extract. In A: polyphenols contents; B: ascorbid acid contents; C: catalase activity; D: superoxide dismutase activity; E: glutathione peroxidase activity. Data are expressed as means±S.D. Different letters means statistically different results ( $p<0.05$ ).

## FIGURE GRAPHICS

Figure 1

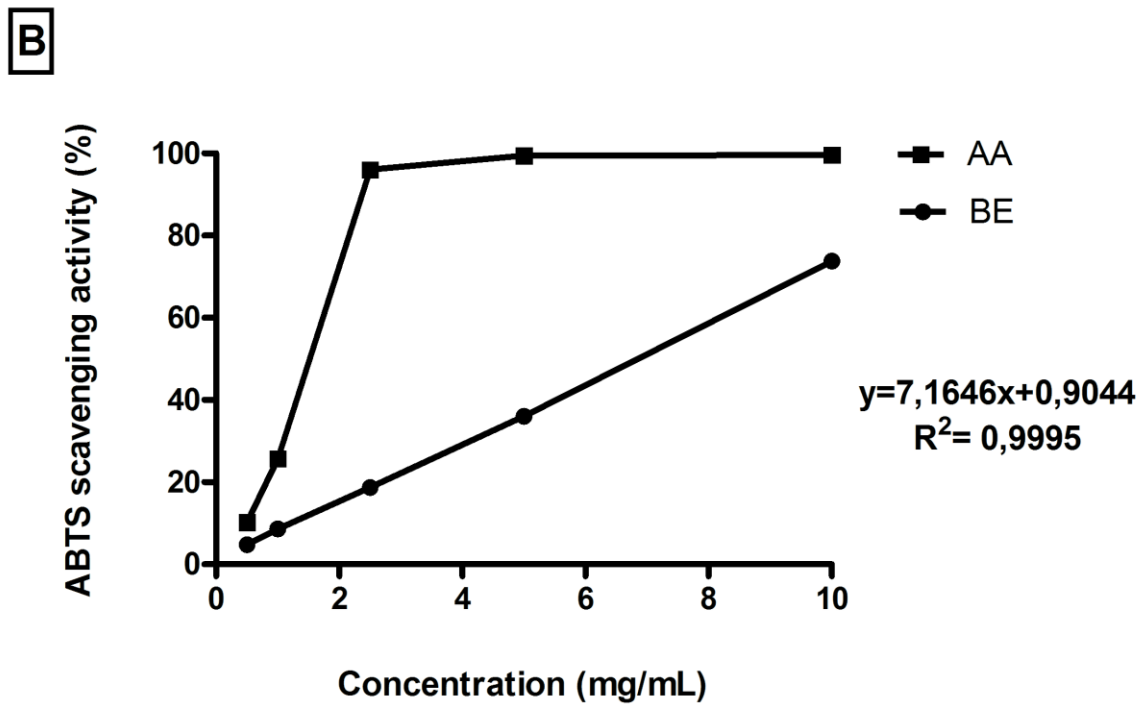
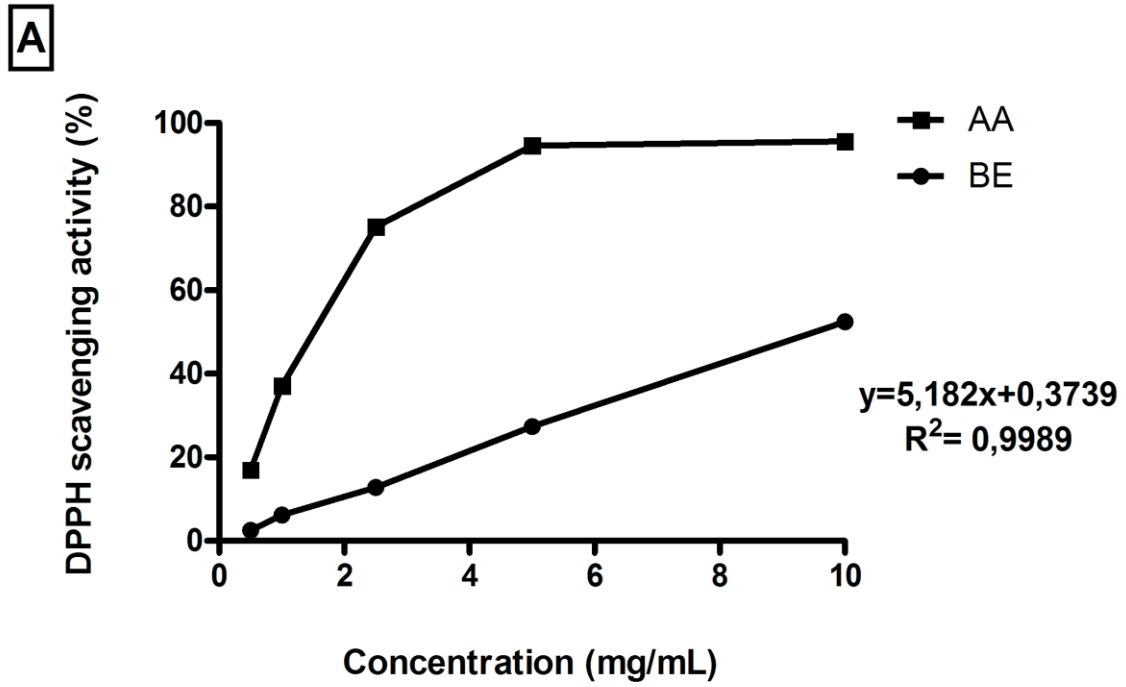


Figure 2

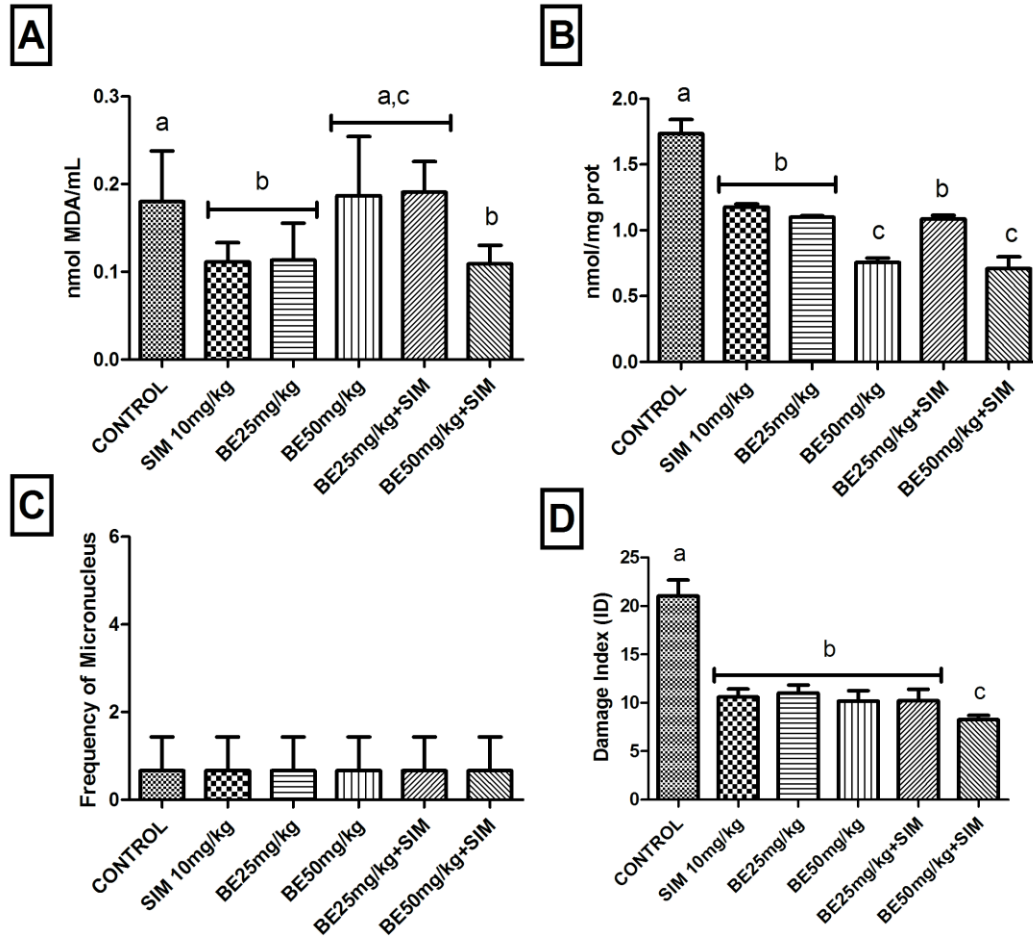
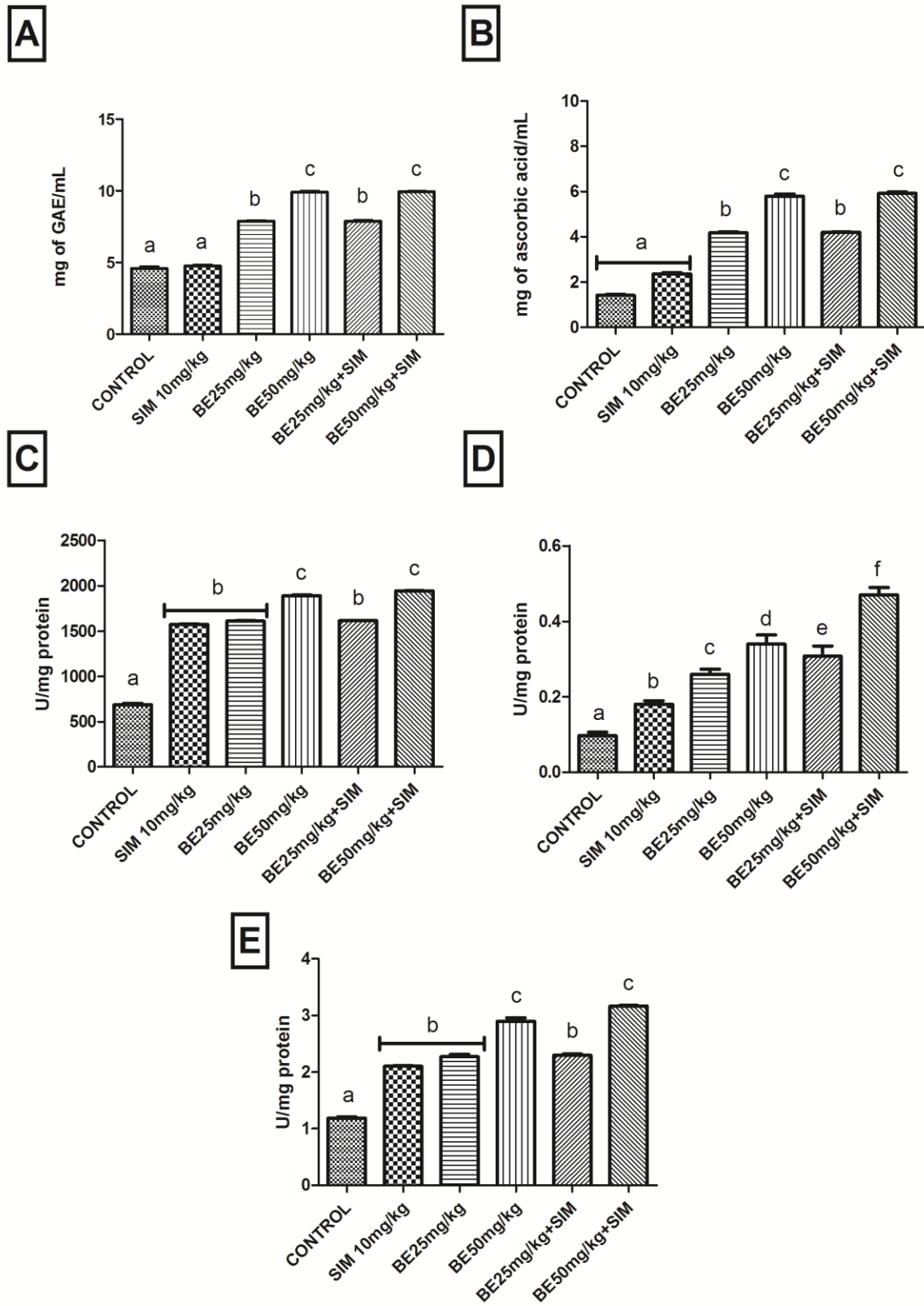


Figure 3





## PARTE III

### 4.0 CONCLUSÃO

De acordo com os resultados apresentados nesta dissertação pode-se inferir que o extrato liofilizado de mirtilo:

- apresenta atividade antioxidante *in vitro*, evidenciada pela atividade sequestradora do radical DPPH e ABTS, o que pode ser atribuído ao seu elevado teor de compostos fenólicos totais (190.620 mg GAE/mL);
- evitou o ganho de peso corporal, diminuiu os níveis plasmáticos de colesterol total, colesterol LDL e triglicérides, bem como aumentou o colesterol HDL. E, este efeito é ligeiramente melhor quando o extrato na dose de 50 mg/Kg está associado à sinvastatina.
- não mostrou efeito significativo sobre o perfil glicêmico;
- diminuiu o espessamento da aorta, mostrando um efeito protetor contra a aterosclerose;
- reduziu significativamente os marcadores de dano cardíaco (CK-total e CK-MB) e homocisteína;
- reduziu significativamente os marcadores de estresse oxidativo, como a peroxidação lipídica e carbonilação de proteínas;
- aumentou significativamente os níveis de polifenóis e ácido ascórbico no plasma;
- diminuiu o dano no DNA de leucócitos;
- aumentou a atividade das enzimas antioxidantes SOD, CAT e GPx, mostrando um importante papel na atividade antioxidante.

Estes resultados mostram que o mirtilo têm efeitos positivos e, portanto, tem potencial para utilização no desenvolvimento de nutracêuticos.

## 5.0 PERSPECTIVAS

Este trabalho tem como perspectivas:

- Medir marcadores inflamatórios como PCR, IL-1B, IL-6, IL-10 e TNF- $\alpha$ ;
- Determinar a concentração das apolipoproteínas (B e A-I);
- Avaliar as principais antocianinas presentes no extrato liofilizado de *Vaccinium ashei* Reade.

## 6.0 REFERÊNCIAS BIBLIOGRÁFICAS

- ABRAHÃO, S. A. **Qualidade da Bebida e Atividade antioxidante do café *in vivo* e *in vitro***. 2007. 92 f. Dissertação (Mestrado em Ciências dos Alimentos). Universidade Federal de Lavras, Lavras, 2007.
- ANTUNES, L. E. C.; MADAIL, J. C. M. Mirtilo: que negócio é esse? **Jornal da Fruta**, n. 159, p. 8, 2005.
- BAGCHI, D. et al. Anti-angiogenic, antioxidant, and anti-carcinogenic properties of a novel anthocyanin-rich berry extract formula. **Biochemistry**, v. 69, n. 1, p. 75-80, 2004.
- BAIGENT, C. et al. Phenolic compounds in plants and agri-industrial by-products: Antioxidant activity, occurrence, and potential uses. **Food Chemistry**, v. 99, n. 1, p. 191-203, 2006.
- BALASUNDRAM, N.; SUNDRAM, K.; SAMMAN, S. Phenolic compounds in plants and agri-industrial by-products: Antioxidant activity, occurrence, and potential uses. **Food Chemistry**, v. 99, p. 191-203, 2006.
- BAÑADOS, M. P. Blueberry production in South America. **ISHS Acta Horticulturae**, n. 715, p. 165-172, 2006.
- BANDYOPADHYAY, D. et al. Oxidative stress-induced ischemic heart disease: protection by antioxidants. **Current Medicinal Chemistry**, v.11, n. 3, p. 369-387, 2004.
- BARBALHO, S. M. et al. *Mentha piperita* effects on wistar rats plasma lipids. **Brazilian Archives of Biology and Technology**, v. 52, n. 5, p. 1137-1143, 2009.
- BASU, A. et al. Blueberries decrease cardiovascular risk factors in obese men and women with metabolic syndrome. **The Journal of Nutrition**, v. 140, n. 9, p. 1582-1587, 2010.
- BECKSTROM, B. W. et al. Correlation between carotid area calcifications and periodontitis: a retrospective study of digital panoramic radiographic findings in pretreatment cancer patients. **Oral Surgery, Oral Medicine, Oral Pathology, Oral Radiology, and Endodontics**, v. 103, n. 3, p. 359-366, 2007.
- BEECHER, G. R. Overview of dietary flavonoids: nomenclature, occurrence and intake. **The Journal of Nutrition**, v. 133, n. 10, p. 3248S-3254S, 2003.
- BELLÓ, A. Dano oxidativo e regulação biológica pelos radicais livres. In: MARRONI, N. P. (Org.). **Estresse Oxidativo e Antioxidantes**. Porto Alegre: Editora Ulbra, 2002.

- BIESALSKI, H. K. Free radical theory of aging. **Current Opinion in Clinical Nutrition and Metabolic Care**, v. 5, n. 1, p. 5-10, 2002.
- BRASIL. Ministério da Saúde. Agência Nacional de Vigilância Sanitária. **Resolução n. 16, 30 de abril de 1999**. Aprova o Regulamento Técnico de Procedimentos para Registro de Alimentos e ou novos ingredientes. Brasília, DF, 1999.
- BROWN, M. S.; GOLDSTEIN, J. L. Scavenging for receptors. **Nature**, v. 343, p. 508-509, 1990.
- BRUCKNER, G. Fatty acids and cardiovascular disease. In: CHOW, C.K. (org.). **Fatty acids in foods and their health implications**. Boca Raton: CRC Press, 2008, p. 1061-1084.
- CASTREJÓN, A. D. R. et al. Phenolic profile and antioxidant activity of highbush blueberry (*Vaccinium corymbosum* L.) during fruit maturation and ripening. **Food Chemistry**, v. 109, n. 3, p. 564–572, 2008.
- CHEN, J. K.; CHOW, S. E. Antioxidants and myocardial ischemia: reperfusion injuries. **Chang Gung Medical Journal**, v. 28, n. 6, p. 369–377, 2005.
- CHO, M. J. et al. Flavonoid glycosides and antioxidant capacity of various blackberry, blueberry and red grape genotypes determined by high-performance liquid chromatography/mass spectrometry. **Journal of the Science of Food and Agriculture**, v. 84, n. 13, p. 1771-1782, 2004.
- CISSE, M. et al. Thermal Degradation Kinetics of Anthocyanins from Blood Orange, Blackberry, and Roselle Using the Arrhenius, Eyring, and Ball Models. **Journal of Agricultural and Food Chemistry**, v. 57, n. 14, p. 6285-6291, 2009.
- COZMA, L. S. The role of antioxidant therapy in cardiovascular disease. **Current Opinion in Lipidology**, v. 15, n. 3, p. 369-371, 2004.
- CUPPARI, L. **Guia de Nutrição Clínica no adulto**. Barueri: Manole, 2002.
- DEFURIA, J. et al. Dietary blueberry attenuates whole-body insulin resistance in high fat-fed mice by reducing adipocyte death and its inflammatory sequelae. **The Journal of Nutrition**, v. 139, n. 8, p. 1510-1516, 2009.
- DELGADO-VARGAS, F.; PAREDES-LÓPEZ, O. **Natural colorants for food and nutraceutical uses**. Boca Raton: CRC Press, 2003.
- DEWICK, P. M. **Medicinal Natural Products: a biosynthetic approach**. 2. ed. West Sussex: Wiley, 2002.

- DULEBOHN, R. V. et al. Effects of blueberry (*Vaccinium ashei*) on DNA damage, lipid peroxidation, and phase II enzyme activities in rats. **Journal of Agricultural and Food Chemistry**, v. 56, n. 24, p. 11700–11706, 2008.
- ECK, P. et al. Blueberry Management In: GALLETTA, G.J.; HIMELRICK, D.G. (org.). **Small fruit crop management**. New Jersey: Prentice Hall, 1990, p. 273-333.
- EMBRAPA/CLIMA TEMPERADO: **Arquivos e Cadernetas de Campo**, 2001.
- ERLUND, I. et al. Consumption of black currant, lingonberries and bilberries increases serum quercetin concentrations. **European Journal of Clinical Nutrition**, v. 57, n. 1, p. 37-42, 2003.
- FACHINELLO, J. C. Mirtilo. **Revista Brasileira de Fruticultura**, v. 30, n. 2, 2008.
- FINAUD, J; LAC, G; FILAIRE, E. Oxidative stress: relationship with exercise and training. **Sports Medicine**, v. 36, n. 4, p. 327-358, 2006.
- FINKEL, T.; HOLBROOK, N. J. Oxidants, oxidative stress and the biology of ageing. **Nature**, v. 408, n. 6809, p. 239-247, 2000.
- FRANCIS, F. J. Anthocyanins and betalains: composition and applications. **Cereal Foods World**, v. 45, n. 5, p. 208-213, 2000.
- GILLHAM, B. et al. Wills: biochemical basis of medicine.. **Oxford: Reed Educational and Professional Publishing Ltd**, v. 3<sup>a</sup> ed., p. 196- 202, 1997.
- GIOVANELLI, G.; BURATTI, S. Comparison of polyphenolic composition and antioxidant activity of wild Italian blueberries and some cultivated varieties. **Food Chemistry**, v. 112, n. 4, p. 903-908, 2009.
- GUTTERIDGE, J., HALLIWELL, B. Free radicals and antioxidants in the year 2000: a historical look to the future. **Annals of the New York Academy of Sciences**, v. 899, n. 1, p. 136-147, 2000.
- HALLIWELL B, GUTTERIDGE J.M.C. **Free Radicals in Biology and Medicine**. 4 ed. New York: Oxford University Press, 2007.
- HALLIWELL, B. et al. The characterization of antioxidants. **Food Chemistry and Toxicology**, v. 33, n. 7, p. 601-617, 1995.
- HALLIWELL, B. Free radicals, antioxidants, and human disease: curiosity, cause, or consequence? **Lancet**, v. 344, n. 8924, p. 721-724, 1994.

- HALLIWELL, B. Reactive Oxygen Species in Living Systems: Source, Biochemistry, and Role in Human Disease. **The American Journal of Medicine**, v. 91, n. 3, p. 14-22, 1991.
- HALLIWELL, B.; GUTTERIDGE, M. **Free Radicals in Biology and Medicine**. 3. ed. New York: Oxford University Press, 1999.
- HALLIWELL, B.; WHITEMAN, M. Measuring reactive species and oxidative damage *in vivo* and in cell culture: how should you do it and what do the results mean? **British Journal of Pharmacology**, v. 142, n. 2, p. 231-255, 2004.
- HUANG, W. Y. et al. Survey of antioxidant capacity and phenolic composition of blueberry, blackberry, and strawberry in Nanjing. **Journal of Zhejiang University Science B**, v. 13, n. 2, p. 94-102, 2012.
- INOUE, N., NAGAO, K.; NOMURA, S.; SHIROUCHI, B.; INAFUKU, M.; HIRABARU, H.; NAKAHARA, N.; NISHIZONO, S.; TANAKA, T.; YANAGITA, T. Effect of *Vaccinium ashei* reade leaf extracts on lipid metabolism in obese OLETF rats. **Bioscience, Biotechnology, and Biochemistry**, v. 75, p. 2304-2308, 2011.
- JACKSON, R. Chemical Constituents of grapes. In:\_\_\_\_\_. (Org.). **Wine science: principles and applications**. London: Academic Press, 1994. p. 178-219.
- JOSEPH, J. A. et al. Reversals of age-related declines in neuronal signal transduction, cognitive, and motor behavioral deficits with blueberry, spinach, or strawberry dietary supplementation. **The Journal of Neuroscience**, v. 19, n. 18, p. 8114–8121, 1999.
- KALEA, A.Z. et al. Vascular reactivity is affected by dietary consumption of wild blueberries in the Sprague-Dawley rat. **Journal of Medicinal Food**, v. 12, n. 1, p. 21-28, 2009.
- KALT, W. et al. Antioxidant capacity, Vitamin C, Phenolics, and Anthocyanins after Fresh Storage of Small Fruits. **Journal of Agricultural and Food Chemistry**, v. 47, n. 11, p. 4638-4644, 1999.
- KAWAMORI, R. et al. Prevalence of carotid atherosclerosis in diabetic patients. Ultrasound high-resolution B-mode imaging on carotid arteries. **Diabetes Care**, v. 15, n. 10, p. 1290-1294, 1992.
- KLUGE, R. A. et al. Frigoconservação de frutos de mirtilo (*Vaccinium ashei* Reade) CV. Climax. **Revista Brasileira de Agrociência**, v. 1, n. 3, p. 185-188, 1995.
- KONCZAK, I.; ZHANG, W. Anthocyanins: more than nature's colours. **Journal of Biomedicine and Biotechnology**, v. 5, p. 239-240, 2004.

- KONG, J. M. et al. Analysis and biological activities of anthocyanins. **Phytochemistry**, v. 64, n. 5, p. 923-933, 2003.
- KOVANEN, P.T.; PENTIKAINEN, M.O. Circulating lipoproteins as proinflammatory and anti-inflammatory particles in atherogenesis. **Current Opinion in Lipidology**, v. 14, n. 5, p. 411-419, 2003.
- LAJOLO, F. M. Alimentos funcionais: uma visão geral. In: ANGELIS, R. C. (Org.). **Importância de Alimentos Vegetais na proteção da saúde: fisiologia da nutrição protetora e preventiva de enfermidades degenerativas**. São Paulo: Atheneu, 2001.
- LEDUC, C. et al. Plants used by the Cree Nation of Eeyou Istchee (Quebec, Canada) for the treatment of diabetes: A novel approach in quantitative ethnobotany. **Journal of Ethnopharmacology**, v. 105, n. 1-2, p. 55-63, 2006.
- LEE, R.T.; LIBBY, P. The unstable atheroma. **Arteriosclerosis, Thrombosis and Vascular Biology**, v. 17, p. 1859-1867, 1997.
- LEE, S. J. et al. Identification of volatile components in basil (*Ocimum basilicum*) and thyme leaves (*Thymes vulgaris* L.) and their antioxidant properties. **Food Chemistry**, v. 91, n. 1, p. 131-137, 2005.
- LI, C. et al. Composition of polyphenols and antioxidant activity of rabbiteye blueberry (*Vaccinium ashei*) in Nanjing. **Journal of Agricultural and Food Chemical**, v. 61, n. 3, p. 523-531, 2013.
- LIMA, V., L., A., G.; GUERRA, N., B. Antocianinas: Atividade Antioxidante e Biodisponibilidade. **Boletim da Sociedade Brasileira de Ciência e Tecnologia de Alimentos**, n. 37, p. 121-128, 2003.
- LOHACHOOMPOL, V. et al. Determination of anthocyanins in various cultivars of highbush and rabbiteye blueberries. **Food Chemistry**, v. 111, n. 1, p. 249-254, 2008.
- LOPEZ, A. D. et al. Global and regional burden of disease and risk factors, 2001: systematic analysis of population health data. **Lancet**, v. 367, n. 9524, p. 1747-1757, 2006.
- LUSIS, A. J. Atherosclerosis. **Nature Reviews**, v. 407, n. 6801, p. 233-241, 2000.
- MAGALHÃES, C.C; CHAGAS, A.C.P.; LUZ, P.L.; Hipertrigliceridemia: implicações clínicas e terapêuticas. **Revista da Sociedade de Cardiologia do Estado de São Paulo**, v. 15, n. 6, p. 483-488, 2005.
- MANACH, A. et al. Polyphenols: food sources and bioavailability. **American Journal of Clinical Nutrition**, v. 79, n. 5, p. 727-747, 2004.

- MANHITA, A. C.; TEIXEIRA, D. M.; COSTA, C. T. Application of sample disruption methods in the extraction of anthocyanins from solid or semi-solid vegetables samples. **Journal of Chromatography A**, v. 1129, n. 1, p. 14-20, 2006.
- MAYNE, S.T. Antioxidant nutrients and chronic disease: use of biomarkers of exposure and oxidative stress status in epidemiologic research. **Journal of Nutrition**, v. 133, n. 3, p. 933S-940S, 2003.
- MOLAN, A.L.; LILA, M.A.; MAWSON, J. Satiety in rats following blueberry extract consumption induced by appetite-suppressing mechanisms unrelated to in vitro or in vivo antioxidant capacity. **Food Chemistry**, v. 107, n. 3, p. 1039-1044, 2008.
- NAVAB, M., ANANTHRAMAIAH G. M., REDDY, S. T., VAN LENTEN, B.J., ANSELL, B.J., FONAROW, G.C., VAHABZADEH, K., HAMA, S., HOUGH, G., et al. The oxidation hypothesis: the role of oxidized phospholipids and HDL. **Journal of Lipid Research**, v. 45, p.993-1007, 2004.
- NETO, C. C. Cranberry and blueberry: evidence for protective effects against cancer and vascular diseases. **Molecular Nutrition & Food Research**, v.51, n. 6, p. 652–664, 2007.
- NOONAN, W. P.; NOONAN, C. Legal requirements for “functional foods” claims, **Toxicology Letters**, v. 150, n. 1, p. 19-24, 2004.
- PAPAHARALAMBUS, C. A., GRIENGLING, K. K. Basic mechanisms of oxidative stress and reactive oxygen species in cardiovascular injury. **Trends in Cardiovascular Medicine**, v. 17, n. 2, p. 48-54, 2007.
- PAPANDREOU, M. A. et al. Effect of a polyphenol-rich wild blueberry extract on cognitive performance of mice, brain antioxidant markers and acetylcholinesterase activity. **Behavioural Brain Research**, v. 198, n. 2, p. 352-358, 2009.
- PICCINATO, C. E.; CHERRI, J.; MORIYA, T. Hipertensão e doença arterial periférica. **Revista Brasileira de Hipertensão**, v. 8, n. 3, p. 306-315, 2001.
- PIMENTEL, C. V. M. B.; FRANCKI, V. M.; GOLLUCKE, A. P. B. **Alimentos funcionais: Introdução às principais substâncias bioativas em alimentos**. São Paulo: Varela, 2005.
- PRIOR, R. L. et al. Purified berry anthocyanins but not whole berries normalize lipid parameters in mice fed an obesogenic high fat diet. **Molecular Nutrition & Food Research**, v. 53, n. 11, 1406–1418, 2009.
- PRIOR, R. L.; CAO, G. Analysis of botanicals and dietary supplements for antioxidant capacity: a review. **Journal of AOAC International**, v. 83, n. 4, p. 950–956, 2000.



- RASEIRA, M. C. B.; ANTUNES, L. E. C. A cultura do mirtilo (*Vaccinium myrtillus*). 1. ed. Pelotas: Embrapa Clima Temperado, 2004.
- RIBEIRO, S. M. R. et al. Antioxidantes na dieta. In: COSTA, N. M. B.; PELUZIO, M. C. G. (Org.). **Nutrição Básica e Metabolismo**. Viçosa: UFV, 2008. p. 235-260.
- RICE-EVAN, C. A.; PACKER, L. **Flavonóides in Health and disease**. 2. ed. London: Marcel Dekker, 2003.
- ROBERFROID, M. Functional food concept and its application to prebiotics. **Digestive and Liver Disease**, v. 34, n. 2, p. S105-S110, 2002.
- ROSENSON, R. S. Statins in atherosclerosis: lipid-lowering agents with antioxidant capabilities. **Atherosclerosis**, v. 173, n. 1, p. 1-12, 2004.
- ROSS, J. A.; KASUM, C. M. Dietary flavonoids: bioavailability, metabolic effects, and safety. **Annual Review of Nutrition**, v. 22, p. 19-34, 2002.
- ROSS, R. Atherosclerosis – an inflammatory disease. **The New England Journal of Medicine**, v. 340, n. 2, p. 115-126, 1999.
- SANTOS, A. M. Situação e perspectiva do mirtilo no Brasil. Embrapa Clima Temperado Pelotas, RS. **Série Documentos**, n. 134, p. 282-285, 2004.
- SCALBERT, A.; WILLIAMSON, G. Dietary intake and bioavailability of polyphenols. **Journal of Nutrition**, v. 130, n. 8, p. 2073-2085, 2000.
- SCHIAVO, M. et al. Influência da dieta na concentração sérica de triglicérides. **Jornal Brasileiro de Patologia e Medicina Laboratorial**, v. 39, n. 4, p. 283-288, 2003.
- SHAHIDI, F.; JANITHA, P. K.; WANASUNDARA, P. D. Phenolic antioxidants. **Critical reviews in food science and nutrition**, v. 32, n. 1, p. 67-103, 1992.
- SHAUGHNESSY, K. S. et al. Diets containing blueberry extract lower blood pressure in spontaneously hypertensive stroke-prone rats. **Nutrition Research**, v. 29, n. 2, p. 130-138, 2009.
- SILVA, S. D. A. et al. Caracterização de genótipos de mirtilo utilizando marcadores moleculares. **Revista Brasileira de Fruticultura**, v. 30, n. 1, p. 180-184, 2008.
- SINGH, U.; JIALAL, I. Oxidative stress and atherosclerosis. **Pathophysiology**, v. 13, p. 129-142, 2006.
- SIQUEIRA, A.F.A.; ABDALLA, D.S.P.; FERREIRA, S.R.G. LDL: da síndrome metabólica à instabilidade da placa aterosclerótica. **Arquivos Brasileiros de Endocrinologia & Metabologia**, v. 50, no 2, p. 334-343, 2006.

- SPEIT, G. et al. A Detection of DNA effects in human cells with comet assay and their relevance for mutagenesis. **Toxicology Letters**, v. 88, n. 1-3, p. 91-98, 1996.
- SPOSITO, A. C. et al. Sociedade Brasileira de Cardiologia. IV Diretriz Brasileira sobre Dislipidemia e Prevenção da Aterosclerose. **Arquivos Brasileiros de Cardiologia**, v. 88, 2014.
- STEINBERG, D. A critical look at the evidence for the oxidation of LDL in atherogenesis. **Atherosclerosis**. v. 131, p. S5-S7, 1997
- STEINBERG, D. The LDL modification hypothesis of atherogenesis: an update. **Journal of Lipid Research**, v. 50, p. S376–S381, 2009.
- STRIK, B. Blueberry: an expanding world crop. **Chronica Horticulturae**, v.45, p.7-12, 2005.
- STULL, A. J. et al. Bioactive in blueberries improve insulin sensitivity in obese, insulin-resistant men and women. **Journal of Nutrition**. v. 140, p. 1764-1768, 2010.
- TAIZ, L.; ZEIGER E. **Fisiologia Vegetal**. 3. ed. Porto Alegre: Editora Artmed, 2004.
- TAKESHITA, M. et al. Proanthocyanidin from blueberry leaves suppresses expression of subgenomic hepatitis C virus RNA. **The Journal of Biological Chemistry**, v. 284, n. 32, p. 21165–21176, 2009.
- VINCENT, H. K.; INNES, K.; VINCENT, K. R. Oxidative stress and potential interventions to reduce oxidative stress in overweight and obesity. **Diabetes, Obesity & Metabolism**, v. 9, n. 6, p. 813-839, 2007.
- WANG, S. Y. et al. Fruit quality, antioxidant capacity, and flavonoid content of organically and conventionally grown blueberries. **Journal of Agricultural and Food Chemistry**, v. 56, n. 14, p. 5788-5794, 2008.
- WANG, Y. et al. Dietary supplementation with blueberries, spinach, or spirulina reduces ischemic brain damage. **Experimental Neurology**, v. 193, n. 1, p. 75–84, 2005.
- WOLFE, K. L. et al. Cellular antioxidant activity of common fruits. **Journal of Agricultural and Food Chemistry**, v. 56, n. 18, p. 8418-8426, 2008.
- WORLD HEALTH ORGANIZATION (WHO). Regional office for the Western Pacific. **The world medicines situation 2011: traditional medicines: global situation, issues and challenges**. Geneva: WHO, 2011. 12p.
- WU, X. et al. Dietary blueberries attenuate atherosclerosis in apolipoprotein E-deficient mice by upregulating antioxidant enzyme expression. **Journal of Nutrition**, v. 140, p. 1628-1632, 2010.

- WU, X. et al. Lipophilic and hydrophilic antioxidant capacities of common foods in the United States. **Journal of Agricultural and Food Chemistry**, v. 52, p. 4026-4037, 2004.
- YI, W. et al. Phenolic compounds from blueberries can inhibit colon cancer cell proliferation and induce apoptosis. **Journal of Agriculture and Food Chemistry**, v. 53, n. 18, p. 7320–7329, 2005.
- ZUANAZZI, J. A.; MONTANHA, J. A. **Farmacognosia: da planta ao medicamento**. 5. ed. Porto Alegre: Editora da UFSC, 2004.

## 7.0 ANEXOS

### Protocolo de aprovação do projeto pelo CEUA-UNIPAMPA



MINISTÉRIO DA EDUCAÇÃO  
FUNDAÇÃO UNIVERSIDADE FEDERAL DO PAMPA  
(Lei nº 11.640, de 11 de janeiro de 2008)

Pró-Reitoria de Pesquisa

#### COMISSÃO DE ÉTICA NO USO DE ANIMAIS - CEUA

Fone: (55) 3413 4321, E-mail: [cena@unipampa.edu.br](mailto:cena@unipampa.edu.br)

---

#### PROTOCOLO Nº 035/2012

**Título:** INVESTIGAÇÃO DE VIAS DE ESTRESSE OXIDATIVO EM RATOS WISTAR HIPERCOLESTEROLÊMICOS SUPLEMENTADOS COM EXTRATO DOS FRUTOS DE *Vaccinium ashei* R. (MIRTILO)

**Pesquisador:** Vanusa Manfredini

**Campus:** Uruguaiana

**Telefone:** (55) 3402 0227

**E-mail:** [vanusamanfredini@unipampa.edu.br](mailto:vanusamanfredini@unipampa.edu.br)

Após a análise detalhada do projeto de pesquisa a relatoria da CEUA-Unipampa emite parecer **FAVORÁVEL** para o cadastro do protocolo e execução do referido projeto.

A handwritten signature in blue ink, appearing to read 'Luiz E. Henkes'.

Luiz E. Henkes  
Professor Adjunto  
Coordenador do CEUA/Unipampa

## Certificado de apresentação do trabalho

# 40<sup>o</sup> Congresso Brasileiro de Análises Clínicas

13<sup>o</sup> Congresso Brasileiro de Citologia Clínica  
4<sup>a</sup> Jornada Latinoamericana de Genética Forense  
16 a 19 de junho de 2013 | Costão do Santinho - Florianópolis - SC

## CERTIFICADO

CERTIFICAMOS que o trabalho

**PERFIL LIPÍDICO E GLICÊMICO DE RATOS WISTAR HIPERCOLESTEROLÊMICOS TRATADOS COM EXTRATO DOS FRUTOS DE VACCINIUM ASHEI R (MIRTILO)**

dos autores **DEISE JAQUELINE STRÖHER, MURIEL PANDO PEREIRA, RITIÉLE PINTO COELHO, ANGÉLICA APARECIDA DA COSTA GÜLLICH, BRUNA COCCO PILAR, JAMILA BENVENEGU BRUNO, MARISTELA WITFEL, LEANDRO LEAL GALARÇA, VANUSA MANFREDINI** foi apresentado na sessão de temas livres do Congresso.

Rio de Janeiro, 19 de junho de 2013

Realização



**SBAC**  
Sociedade Brasileira de Análises Clínicas

Entidades Parceiras




**SLAGF**  
Sociedade Latinoamericana de GENÉTICA FORENSE



Dr. Irineu Keiserman Grinberg  
Presidente da Sociedade Brasileira de Análises Clínicas



Dr. Carlos Eduardo de Queiroz Lima  
Presidente da Sociedade Brasileira de Citologia Clínica



Dra. Maria Elizabeth Menezes  
Presidente da Sociedade Latinoamericana de Genética Forense e Presidente da Comissão Científica do 40<sup>o</sup> CBAC



Dr Tércio Egor Paulo Kasten  
Presidente do 40<sup>o</sup> Congresso Brasileiro de Análises Clínicas